



## Review Article

# Gluten is a Proinflammatory Inducer of Autoimmunity



Aaron Lerner<sup>1,2\*</sup>, Carina Benzvi<sup>1</sup> and Aristo Vojdani<sup>3</sup>

<sup>1</sup>Chaim Sheba Medical Center, The Zabludowicz Research Center for Autoimmune Diseases, Tel Hashomer, Israel; <sup>2</sup>Ariel Campus, Ariel University, Ariel, Israel; <sup>3</sup>Immunosciences Lab., Inc., Los Angeles, CA, USA

Received: August 26, 2023 | Revised: December 09, 2023 | Accepted: April 30, 2024 | Published online: June 28, 2024

## Abstract

Gluten has multiple harmful effects that compromise human health, not only in gluten-dependent diseases but also in non-gluten-affected chronic inflammatory conditions. After consumption, the indigestible gluten peptides are modified by luminal microbial transglutaminase or transported through the gut epithelium to interact with the highly populated mucosal immune cells. As a disruptor of gut permeability, gluten peptides compromise tight junction integrity, allowing foreign immunogenic molecules to reach internal compartments. Gliadin peptides are distributed systemically to remote organs, where they encounter endogenous tissue transglutaminase. Following post-translational deamidation or transamidation, the peptides become immunogenic and pro-inflammatory, inducing organ dysfunction and pathology. Cross-reactivity and sequence homology between gluten/gliadin peptides and human epitopes may contribute to molecular mimicry in autoimmunity induction. A gluten-free diet can prevent these phenomena through various mechanisms. As proof of concept, gluten withdrawal alleviates disease activity in chronic inflammatory, metabolic, and autoimmune conditions, and even in neurodegeneration. We recommend combining the gluten-free and Mediterranean diets to leverage the advantages of both. Before recommending gluten withdrawal for non-gluten-dependent conditions, patients should be asked about gut symptomatology and screened for celiac-associated antibodies. The current list of gluten-induced diseases includes celiac disease, dermatitis herpetiformis, gluten ataxia, gluten allergy, and non-celiac gluten sensitivity. In view of gluten being a universal pro-inflammatory molecule, other non-celiac autoinflammatory and neurodegenerative conditions should be investigated for potential gluten avoidance.

## Introduction

Inflammation is a vital biological response that regulates interactions between humans and the environment, with nutrition playing a crucial role. Due to the surge in chronic inflammatory diseases,<sup>1</sup> and increasing interest in anti-inflammatory dietary therapy,<sup>2,3</sup> the exploration of pro-inflammatory nutrients has become a primary focus for clinical and scientific communities.<sup>4</sup> In fact, the understanding of immune system-driven chronic inflammation and its associated chronic diseases are still not well-developed. The contribution of dietary constituents to inflammatory, metabolic, autoimmune, cancerous, allergic, and neurodegenerative diseases remains poorly defined. The frequently consumed Western diet is considered pro-inflammatory,<sup>5</sup> while vegetarian, non-processed, and traditional foods are recommended as anti-inflammatory.<sup>6,7</sup>

**Keywords:** Pro-inflammatory nutrients; Anti-inflammatory nutrients; gluten; Gliadins; Celiac disease; Autoimmune diseases; Chronic inflammation.

**\*Correspondence to:** Aaron Lerner, Chaim Sheba Medical Center, The Zabludowicz Research Center for Autoimmune Diseases, Tel Hashomer 5262000, Israel. ORCID: <https://orcid.org/0000-0002-6779-4090>. Tel: +972-525919484, E-mail: aaronlerner1948@gmail.com

**How to cite this article:** Lerner A, Benzvi C, Vojdani A. Gluten is a Proinflammatory Inducer of Autoimmunity. *J Transl Gastroenterol* 2024;2(2):109–124. doi: 10.14218/JTG.2023.00060.

Since it is impossible to cover all pro-inflammatory nutrients, this review will focus on the role of gluten/gliadin in celiac disease (CD)-induced inflammation, and explore their potential involvement in other non-celiac chronic inflammatory conditions. Gluten is composed of two main proteins: glutenin and gliadin. Gliadins make up about 70% of the protein in gluten and are the molecules responsible for the harmful immune response that results in intestinal injury in CD. Since the gut is the entry point for gluten and a crossroads for multiple nutrients, food additives, microbes, enzymatic digestion, and absorption, various gluten-affected luminal events irradiate peripherally, inducing remote organ, gluten-related, inflammatory damage.<sup>8,9</sup> The luminal content impacts the enteric ecosystem. Certain dietary components, like gluten, breach tight junction integrity, resulting in increased intestinal permeability, and induce changes in the composition and diversity of the microbiome towards disease-specific dysbiosis or pathobiosis. Finally, the enhanced local enzymatic capacity for post-translational modification of proteins can turn naïve peptides to lose their tolerance and become auto-immunogenic ones. The present narrative review expands on the multiple gut-originated axes and their relationship to remote organ autoimmune diseases. Brain, joint, bone, endocrine, liver, kidney, heart, lung, and skin autoimmune diseases are connected to the deregulated events in the intestinal luminal compartment, forming the gut-systemic organ axe. Be-

ing a universal pro-inflammatory protein, affecting multiple body compartments, organs and tissue and systemically distributed, gluten peptides should be thoroughly investigated for their potential detrimental effects. If substantiated, traditional gluten dependent diseases should be ruled out and gluten-free Mediterranean diets should be recommended.

#### **Gluten-induced inflammation in celiac disease**

The incidence of celiac disease is around 1–2% of the global population, representing a chronic, autoimmune, multisystem, inflammatory, and immune-mediated condition.<sup>10</sup> The intestine is the primary target organ; however, extra-intestinal organs are affected as well.<sup>8,9</sup> Currently, the only accepted and proven nutrient that induces the disease is gluten, a general name for proteins found in many grains, such as wheat, barley, rye, and partially in oats.<sup>11</sup> Interestingly, “gluten” stands for “glue” in Latin, named for its adhesive and viscoelastic properties. The autoantigen associated with CD is tissue transglutaminase (tTG),<sup>12,13</sup> and several antibodies are used for its serological diagnosis.<sup>14–16</sup>

Celiac disease fulfills the criteria of a gluten-induced autoimmune inflammatory condition in the following aspects<sup>17–19</sup>: histologically, there is epithelial and mucosal inflammatory destruction, presented by villous atrophy and intraepithelial lymphocytosis<sup>20,21</sup>; immunologically, the adaptive<sup>22</sup> and innate immune<sup>23</sup> systems are activated<sup>24,25</sup>; there is a surge in pro-inflammatory cytokines,<sup>26</sup> where gliadin peptides induce increased levels of IL-15, IFN-γ, IL-6, tumor necrosis factor (TNF)-α, IL-1β, CCL2, CCL3, and many more<sup>27</sup>; dysbiosis, a feature of many autoimmune diseases (ADs), exists in the celiac gut lumen, and gliadin can directly induce intestinal flora dysbiosis.<sup>28–30</sup> Upon successful gluten withdrawal, all the above-mentioned inflammatory features are significantly ameliorated<sup>31,32</sup>; refractory CD and celiac crisis respond to steroids or immunosuppressive therapy<sup>33</sup>; and finally, genetic predisposition is essential for the disease’s development.<sup>25</sup> It can be summarized that CD is a hallmark condition where genetics, inflammation, autoimmunity, dysbiosis, and environmental gluten intersect.<sup>25,34–38</sup> It is well accepted that gluten-containing cereals contribute to chronic inflammatory conditions and various ADs, primarily by inducing gut dysbiosis, enhancing intestinal permeability, and initiating a pro-inflammatory immune response.<sup>6,39–42</sup> Interestingly, in addition to gluten, other wheat components, like lectins and enolase, might also act as proinflammatory molecules.<sup>43–46</sup>

#### **The place of gluten in human nutrition and as a food additive**

Wheat is a major crop grown in most countries. Its annual production is  $7.34 \times 10^8$  tons, cultivated on an area of  $2.14 \times 10^6$  km<sup>2</sup>, which sums up to the size of Greenland.<sup>47</sup>

Wheat consumption surpasses all other crops combined, making it the world’s most favored staple food. The annual increase in gluten usage as a processed food additive over the last four to six decades is estimated at  $1.8 \pm 0.4\%$ .<sup>6</sup> The global vital wheat gluten market, valued at \$2 billion in 2019, is projected to reach \$2.74 billion by 2027, representing a compound annual growth rate of 4% during the forecast period.<sup>48</sup>

Discovered in the Fertile Crescent nearly 14,000 years ago,<sup>49</sup> wheat-based foods became a staple after domestication. Wheat is a major source of protein, with gluten making up 80% of its total protein content.<sup>6,50</sup>

Gluten is a protein naturally found in certain prolamins, including wheat, barley, and rye. Although oats are considered gluten-free, they are often cross-contaminated with gluten.<sup>11,51</sup> Besides prolamins, gluten is found in numerous non-nutritional products,

such as medications, toothpaste, and cosmetics. It acts as a binder, holding substances together and adding a “stretchy” quality. When omitted in baking, the resultant dough tears easily.<sup>52</sup>

In Western countries, wheat contributes significantly to health, providing dietary fiber, B vitamins, and mineral micronutrients, notably selenium, iron, and zinc.<sup>53</sup> There are two perspectives on wheat and gluten: public perception and reality.<sup>50,53</sup> Recently, the roles and functions of wheat and gluten have been scrutinized in unproven or pseudoscientific publications and popular media reports, giving the impression that wheat or gluten consumption has a deleterious and addictive effect on human health.<sup>50,53</sup> The common claim that gluten-free foods are inherently healthy is not well-supported by scientifically accepted controlled studies. Consequently, wheat/gluten withdrawal fashionistas have impacted multiple Western societies, changing their dietary habits.<sup>50,53</sup> While gluten itself provides no essential nutrients and its consumption can be avoided without compromising human well-being.<sup>54</sup> However, the only scientifically defined indication to withdraw gluten is in well-proven gluten-dependent conditions, namely, CD, dermatitis herpetiformis, gluten ataxia, and gluten/wheat allergy.<sup>31,32,41,53,55–57</sup> Despite the significant interest in non-celiac gluten sensitivity, much remains to be explored about its pathogenesis, and its potential nutritional triggers are still controversial.<sup>55,58–61</sup> A combined gluten-free and Mediterranean diet might be an attractive alternative to address the unhealthy aspects of a gluten-free diet (GFD).<sup>53,62–64</sup>

#### **The side effects of gluten**

The more gluten is explored, the more side effects are disclosed. This topic has recently been reviewed by several groups.<sup>6,8,9,41,42,65–67</sup> There are various adverse effects of gluten that might impact health. These harmful effects are delivered through immunological and toxic pathways, leading to gut dysfunction or inadequacy.

#### **Pro-inflammatory**

As mentioned above, gluten acts as a pro-inflammatory molecule.<sup>8,9,25–27,34–39,68,69</sup> The gliadin p31-43 peptide induces cellular stress, activates proliferative mechanisms of cryptic epithelial cells, drives enterocyte stress, induces a Ca<sup>2+</sup> surge (thus activating the CD autoantigen tTG), triggers a local pro-inflammatory storm, activates the NFκB signaling pathway, inhibits CFTR (cystic fibrosis transmembrane conductance regulator, an ion channel protein), alters vesicular trafficking, activates the inflammasome platform, and reduces autophagy.<sup>68,70,71</sup> Gliadin peptides are pro-oxidative, induce DNA damage, and are pro-apoptotic in in-vitro and ex-vivo studies.<sup>72</sup>

#### **Alters the gut microbiome and increases intestinal permeability**

Gluten decreases the microbiome/dysbiome ratio composition and diversity, suppressing the beneficial metabolome toward inflammation.<sup>28–30,73–77</sup> Intestinal tight junction functional integrity is one of the most conserved protective mechanisms for human survival and is crucial for maintaining intestinal homeostasis. When disrupted, foreign molecules enter the epithelial barrier, come into contact with the subepithelial dense immune systems, and initiate chronic inflammation and autoimmunity. Increased intestinal permeability is a common feature of many of these conditions, including CD,<sup>6,35–38,41,65–68,74,75</sup> where gluten is a major disruptor of tight junction protective function.<sup>6,78–81</sup> Several observations strengthen the gluten-zonulin-increased permeability axis. Zonulin is a meas-

urable blood protein that reflects tight junction functionality. Increased zonulin blood level is considered a marker of an impaired intestinal barrier.<sup>6,78–81</sup> This active axis has been shown to operate in active and non-active CD patients and even in normal controls.<sup>82</sup> Larazotide acetate (AT-1001), a small peptide derived from Vibrio cholerae toxin, is one of the potential non-nutritional pharmacological strategies to treat CD patients.<sup>56</sup> Acting as a modulator of tight junction integrity with its anti-zonulin activity, it has been shown to be superior to placebo in improving gut symptoms in active CD patients.<sup>83</sup> The drug shows promise for counteracting enhanced intestinal permeability in some other chronic systemic diseases,<sup>84–86</sup> including severe COVID-19,<sup>87,88</sup> although it is not yet approved.

### Immunogenicity

One of the major unwanted effects of gluten is its immunological impact, closely connected to the inflammatory response. Gluten is an immunogenic protein that elicits anti-gluten/gliadin antibodies, even in non-CD patients and normal controls.<sup>89–93</sup> In addition, microbial transglutaminase (mTG)-treated gluten peptides in patients with CD are immunogenic.<sup>94</sup> When gliadin is cross-linked to tissue or mTG, transforming the naïve protein into an immunogenic one, CD patients mount substantial levels of neo-epitope tTG and mTG antibodies, respectively.<sup>8,9,14–16,41,66,94–99</sup> Intriguingly, in the presence of tTG and mTG-assisted gliadin docking, gluten/gliadin loses its human body tolerance, resulting in corresponding antibody secretion, representing a classical post-translation modification of proteins.<sup>29,30,100</sup> When tested in CD patients, the neo-epitope fTG and mTG exhibit higher immunogenic activity compared to gliadin undocked enzymes. Furthermore, the tTG neo-epitope IgA+IgG isotypes show higher optical density activity, better reflect intestinal injury, and expose higher specificity and sensitivity by targeting different autoantigens compared to the conventional tTG isotypes.<sup>98,99</sup> The same was found for the mTG neo-epitope.<sup>14,16,95</sup> However, the list of gluten's immunological adverse effects is much more extensive.<sup>41</sup>

*In vitro* studies have shown that gluten induces macrophages to produce proinflammatory cytokines and nitric oxide (NO).<sup>101–103</sup> Upregulated MHCII, co-stimulatory molecules, TRLs, cytokine, and chemokine production were observed in dendritic cells.<sup>104,105</sup> Higher expression of NKG2D and CD71 on NKp46(+) cells has been shown in lymphoid organs.<sup>106</sup> Finally, increased permeability and the production of TNF $\alpha$  and IL-1 $\beta$  were detected when gluten was applied to the Caco-2 cell line.<sup>107</sup>

*In vivo* studies on rats and mice have shown the following compared to controls: cytokine surge in TH1 intestinal and mesenteric lymph nodes, TH1 cytokine pattern in islet infiltrate, and increased number of intestinal pathogenic intraepithelial cells.<sup>108–110</sup> Studies on non-obese diabetic (NOD) mice revealed increased activated intestinal CD4 $^{+}$  T cells, changes in TH1/TH2 intestinal cytokine ratios associated with activated dendritic and TH17 cells, increased natural killer cell cytotoxicity, and cytokine secretion of IFN- $\gamma$  and IL-6.<sup>106,111–113</sup> NKG2D is a proinflammatory, auto-immunogenic co-stimulatory molecule; it is an activating receptor mostly expressed on cells of the cytotoxic arm of the immune system. Gluten withdrawal lowered NKG2D and its ligand expression in NOD and BALB/c mice, attesting to gluten's impact on the co-stimulatory interplay between tolerance and immune inflammation.<sup>114</sup>

When exposed to gliadin/gluten, BALB/c mice showed proportional changes in regulatory T-cell subsets, increased numbers of TH17 in peripheral lymph nodes, proinflammatory cytokine patterns in FOX3 $^{-}$  and FOXP3 $^{+}$  T cells, and robust activation of innate

immune and TH17 cells.<sup>115–117</sup> *Ex vivo* and mice studies showed gluten-induced dendritic cells' production of IL-1 $\beta$  and, interestingly, enhanced neutrophil migration towards gliadin peptides.<sup>118,119</sup>

### Cellular dysfunction and cytotoxicity

At the cellular level, gliadin has been found to drive cytotoxicity, decrease cell viability and differentiation, induce LDH secretion, promote apoptosis, and decrease RNA, DNA, and glycoprotein synthesis when applied to HCT116 cells.<sup>41</sup> In 1976, Hudson *et al.*<sup>120</sup> documented growth inhibition and phenotypic changes in various human cell lines induced by gliadin exposure. Several *in vitro* studies point to the cytotoxicity of this molecule. Gliadin induced agglutination in K562 cells, decreased F-actin content in enteric 407 cell lines, suppressed cell growth and viability, induced apoptosis, and altered redox equilibrium in Caco-2 cells and cell morphology in LoVo, two- and three-dimensional cell culture while causing rearrangement of the cytoskeleton through the zonulin molecular structure. This results in the loss of tight junction functionality in IEC-6 cells.<sup>121</sup>

### Disturbance of oxidative equilibrium

Oxidative equilibrium plays an essential role in cell homeostasis, and its imbalance is involved in many chronic inflammatory diseases. Gliadin-induced oxidative stress was reported extensively on various cell lines, including Caco-2, HT29, SH-SY5Y, T84, and LoVo, and reviewed in depth.<sup>121–123</sup> For example, the content of glutathione was reduced (~20% vs. controls), and the activity of related enzymes was inhibited.<sup>121</sup> The dysfunctional antioxidant machinery can result in inflamed CD intestinal mucosa, making it more vulnerable to further oxidative stress and hindering mucosal recovery.<sup>123</sup>

### Induce apoptosis

Intestinal homeostasis relies heavily on enterocyte viability and death to maintain the high cellular turnover necessary to cope with a hostile environment. Programmed cell death is pivotal for this equilibrium but can be detrimental in pathological conditions. The apoptotic pathway is over-activated in CD patients and plays a key role in inducing gut inflammation.<sup>73,124</sup> Inflammatory response and enteric damage induced by gliadin p31-43 drive multiple programmed cell death pathways in the small intestine of mice.<sup>124</sup>

### Gluten impacts epigenetics

The HLA-DQ2 and HLA-DQ8 haplotypes are widely associated with CD, but some people without these genes still develop the disease. Genetic predisposition can be regulated or affected by epigenetic modifications and cannot account for all reported CD cases. Environmental epigenetics adds substantial understanding to the disease's evolution and its multi-faceted phenotypic presentations.<sup>8,125,126</sup> The main epigenetic pathways include histone modifications, DNA methylation, non-coding RNAs, and RNA methylation, where microRNAs might be used to characterize various classes of CD patients.<sup>125</sup> Gluten affects gene expression by changing methylation status.<sup>122,127</sup> The impact of gliadin on epigenetics has been observed in CD and non-CD MH-SY4Y and Caco-2 cell lines.<sup>122</sup> Wheat-derived peptide epigenetic alterations might be important during the postnatal nutritional transition from maternal breastfeeding or infant formula to complementary gluten consumption.<sup>122,127</sup>

### Gluten affects cellular metabolism

Being pro-inflammatory, cytotoxic, oxidative, apoptotic, and highly immunogenic, gluten peptides can alter fundamental cellular metabolic networks. Wheat-derived peptides induce 50% inhibi-

tion in cellular proliferation, 20% suppression of colony-forming ability, and significantly lower alkaline phosphatase activity during Caco-2 cell line differentiation.<sup>128</sup> Moreover, the peptic-tryptic digestion of wheat inhibited more than half of DNA and RNA synthesis, glycoprotein synthesis, and altered mitochondrial functions in Caco-2 cells.<sup>129,130</sup> Wheat and gluten peptides are important in nutrigenomics and nutrigenetics, revealing various interplays between diet, specific nutritional components, and gene expression.<sup>127</sup>

### **Gluten affects mental health**

A plethora of peripheral and central neurological manifestations affect the celiac population<sup>65,131–135</sup> indicating that gluten consumption can also impact psychiatric behavior and mental health. Anti-neuronal antibodies such as transglutaminase 6, GAD-65, GAD-67, cerebellar peptide, and myelin-associated glycoprotein are part of the CD-associated autoantibodies.<sup>136</sup> During intestinal digestion, resulting gluten fragments have strong opioid activity.<sup>137</sup> These morphine-like substances, called gluten exorphins, have proven opioid effects that might affect mental health.<sup>138</sup> Opioid receptors are scattered throughout the body, including in the gut, brain, and peripheral nervous system. Facing intestinal and blood-brain barrier disruption<sup>139,140</sup> caused by microbes, stress, dietary components, pollutants, alcohol, or over-the-counter drugs, gluten-originated exorphins can impact mental functions.<sup>6,41,42,65,66,131,137</sup> Cognitive impairment and “brain fog” might be associated with CD,<sup>141,142</sup> and responsiveness to gluten withdrawal has been reported.<sup>143–145</sup>

In a more holistic view, the association between oxidative stress, gene expression, dysbiome and its microbiome, impaired gut and brain permeability<sup>146</sup> and gut inflammation associated with gluten-derived peptides, is interrelated and interconnected during the autoimmune cascade evolution in CD.

### **Gluten peptides are systemically distributed**

Prolamins containing gluten are main nutritional staples, and processed food gluten is heavily consumed.<sup>6,41,42,65,66,94</sup> Consequently, gluten is widespread in the environment, in the gut lumen, and in contact with the epithelial monolayer and mucosal immune systems. CD is considered a gradually developing, mostly hyposymptomatic or even asymptomatic chronic enteric inflammatory condition. In reality, it can abruptly erupt as an acute, symptomatic, sometimes life-threatening event involving the gut and extra-intestinal peripheral organs.<sup>8,147</sup> Many *in vivo/ex vivo* or *in vitro* models involving CD duodenal biopsies, intestinal cell lines, or incidental gluten intake have reported acute effects within 48 h of incubation or ingestion, which were recently summarized. These acute phenotypic, cellular, and laboratory events demonstrate the potential ability of gluten peptides to impact the entire human body.<sup>8,9,147,148</sup> A major question is whether gluten/gliadin peptides pass the protective mechanical or immunological intestinal barriers to penetrate inside the body and reach remote compartments and organs. Several observations support the systemic distribution of these peptides, and suggested mechanisms include:

#### **Transepithelial passage of gluten peptides**

The discussion on how gliadin peptides pass the gut epithelial monolayer is ongoing, but it is known that both paracellular and transcellular pathways are involved.<sup>67</sup> There are three methods of transporting molecules through a cell: endocytosis,<sup>67,149</sup> endoplasmic reticulum-assisted transcytosis,<sup>150</sup> and secretory IgA-transferin receptor-assisted translocation of intact gluten peptides below

the epithelium.<sup>151</sup> Paracellularly, following gliadin digest binding to its CXCR3 receptor, increased zonulin levels compromise tight junction function by activating the EGFR-PAR2-MyD88-mediated signaling pathways, resulting in increased intestinal permeability.<sup>152</sup>

Most recently, Stricker S. *et al.* visualized gliadin peptide transport into CD enterocytes using intestinal biopsies and the RACE (Rapid uptake of Antigen into the Cytosol of Enterocytes) cell line.<sup>150</sup> The nutrition-originated peptides were transported through the endoplasmic reticulum and deposited below the enterocyte monolayer. This deposition proves that luminal gluten peptides penetrate the epithelial barrier, hence, facing the mucosal and submucosal immune networks.

In fact, gluten-dependent subepithelial deposits involving IgA-tTG are among the hallmark markers for early CD, even in seronegative patients and before histological damage occurs.<sup>153</sup> The cohabitation of gluten peptides with these specific IgA-tTG deposits in the subepithelial space reinforces the transepithelial transport of gluten peptides.<sup>13,154</sup> Additionally, the immunogenic CD supramolecule, a 33-residue peptide from alpha-2 gliadin, was directly visualized in gluten-sensitive macaques,<sup>155</sup> and gluten-stimulated CD-specific enteric T cells were shown to increase the transepithelial flux of gluten peptides.<sup>156</sup> TTG-gluten polymeric complexes are potent antigens for tTG-specific mucosal B cells, supported by diverse subepithelial gluten-specific T cells.<sup>157</sup> Finally, gluten peptides can be presented by subepithelial dendritic cells.<sup>158</sup> Thus, isolated or complexed gluten/gliadin peptides located in the lamina propria are presented by local antigen-presenting cells, activating the adaptive and innate mucosal networks and inducing CD-specific autoantibodies.

#### **Gluten metabolites are found in human body fluids**

Physio-anatomical logic indicates that urinary secretion of a peptide most likely originates from the bloodstream. Recently, Upadhyay D *et al.* reported that gluten sensitivity expresses itself in a potential CD at the metabolic level before any intestinal damage.<sup>159</sup> Decreased levels of histidine, tyrosine, glycine, and tryptophan, and altered levels of another six metabolites were detected in the mucosa or plasma of potential CD patients compared to active CD patients and healthy controls. Intriguingly, raising the topic of gluten addiction and mental health, gluten metabolites, namely, exorphin B4 and B5, are found in normal human blood.<sup>160,161</sup> A hypothetical mechanism for gluten masking its own toxicity by these gluten-originated exorphins has been suggested.<sup>138</sup> Additionally, multiple gluten-dependent circulating miRNAs that appear before IgA-tTG positivity and are responsive to gluten withdrawal have been characterized.<sup>162,163</sup>

Urinary gluten metabolites have been extensively reported. Gluten dose escalation, gluten-free diet adherence assessment, urinary gluten intake-dependent miRNAs, urine peptidomics analysis, and urinary metabolic alterations have all been documented.<sup>159,163–166</sup>

#### **Tissue transglutaminase whole body distribution and biological functions**

Before describing gluten peptide distribution in tissues and organs, it is important to remember that tTG is ubiquitous in the human body.<sup>13</sup> It is the autoantigen in CD,<sup>12</sup> its prime substrate is gluten,<sup>94</sup> and the enzyme induces posttranslational modification of gluten,<sup>29</sup> making gluten immunogenic in several gluten-dependent conditions.<sup>13,66</sup> Cellular-wise, the enzyme spans all intracellular organelles and compartments, including transmembrane areas, and is secreted extracellularly.<sup>13,167</sup> Tissue-wise, it is ubiquitously ex-

pressed in most human tissues.<sup>13,167</sup> Due to its enzymatic biochemical activities, tTG is involved in multiple human biological events and diseases.<sup>13,167,168</sup> Since gluten peptides circulate systemically, the chances of tTG encountering them are high. The missing part of the tTG-blood-gluten triangle is the localization of gluten/gliadin metabolites in remote extra-intestinal organs.

#### **Gluten metabolites are found in human organs**

Because CD is a multifaceted condition with a plethora of extra-intestinal phenotypes, patients with the disease are at risk of developing remote organ pathologies.<sup>8,9,30,41,42,55,65,66</sup> The enzyme tTG can cross-link numerous protein substrates, and the resulting aggregates can be deposited in various organs.<sup>169</sup> Below are the main organs where tTG and gluten might orchestrate or be involved in local pathologies:

#### **Gluten deposits in the cerebellum**

Gluten ataxia is an autoimmune ataxia and an integral part of gluten-dependent ADs. Brain IgA-tTG2 and tTG6 deposits have been reported in patients, primarily in the cerebellum, pons, and medulla.<sup>170,171</sup> Intense perivascular deposition and inflammation might allow circulating gluten peptide and IgA-tTG2/tTG6 antibody entry through a leaky blood-brain barrier, depositing in the central nervous system.<sup>172</sup> Indeed, when serum from gluten ataxia patients was injected into the ventricles of mice, ataxia developed within 3 h post-injection.<sup>172,173</sup> Moreover, cross-reactive antibodies are shared between gluten peptides and Purkinje cell epitopes, suggesting a potential molecular mimicry pathway to cerebellar autoimmunogenesis.<sup>174</sup> Despite these findings, the characterization of gluten-IgA-tTG2/tTG6 deposits in the patient's cerebellum requires further evaluation.

#### **Gluten peptides impact chronic inflammatory brain conditions**

The gut-brain axis was recently described,<sup>131,135,175,176</sup> but the role of nutrients in activating these pathways is not clearly defined.<sup>6,8,9,29,30,177</sup> Recent papers strengthen the potential relationship between gluten consumption and neurodegenerative, neuroinflammatory, and central ADs in susceptible individuals.<sup>65,66,134,178</sup> It appears that gluten peptides contribute to neurodegeneration and chronic brain inflammatory diseases.<sup>65,66,178</sup> By specific dysbiosis, enhanced gut permeability, many cross-reactive antibodies with sequence similarity to human brain epitopes, and multiple adverse effects described above, the circulating repertoire of gluten peptides is involved in neurodegeneration.<sup>65,66,134</sup> The combined cross-reactivity and sequence similarity suggest molecular mimicry and allude to autoimmune mechanisms resulting in gluten-related brain conditions.

#### **Gluten deposit in the thyroid**

Hashimoto's thyroiditis and CD often overlap, sharing genetic, environmental, symptomatic, immunogenic, pathological, hormonal, and even serological aspects.<sup>179</sup> Both entities are integral parts of polyendocrinopathy syndrome.<sup>180</sup> There is a continuous debate concerning whether and when to screen patients for CD serology and if there is a place for a gluten-free diet in autoimmune thyroid diseases.<sup>179,181–183</sup> The tTG enzyme resides in the thyroid follicles and extracellular matrix.<sup>184,185</sup> In CD patients, tTG antibodies bind to these thyroid regions, and their levels correlate with thyroid peroxidase antibody activity.<sup>185</sup> The findings suggest that CD-associated antibodies could be involved in thyroid dysfunction, thus reinforcing the gut-thyroid axis. Supporting this, Vojdani *et al.* applied affinity-purified antibodies made against wheat, al-

pha-gliadin peptide, and wheat germ agglutinin to various human tissue antigens, finding moderate to strong reactions with thyroid peroxidase and many other autoantigens.<sup>44,186</sup>

#### **Gluten deposits in the skin**

Dermatitis herpetiformis is a dermatological gluten-dependent disease characterized by cutaneous anti-tTG3 IgA deposits, where tTG3c is the autoantigen of the disease.<sup>187</sup> Interestingly, similar aggregates can be detected in the skin of CD patients.<sup>188</sup> tTG3 is a member of the tTG family and can cross-link with its favorable gluten/gliadin peptides. The skin anti-tTG3 IgA deposits in dermatitis herpetiformis mirror the subepithelial IgA-tTG deposits in CD. The herpetiform dermatological eruption is only one of many gluten-dependent autoimmune manifestations of the skin.<sup>189</sup>

#### **Gluten deposits in the pancreas**

Type 1 diabetes mellitus is highly associated with CD,<sup>8,136,180,190</sup> but the role of gluten in inducing insulitis remains controversial.<sup>191</sup> Some researchers claim that gluten-containing cereals are associated with an increased risk of pancreatic islet autoimmunity,<sup>79,192–194</sup> while others see no such connection.<sup>195</sup> Notably, gluten peptides have been shown to localize in the pancreatic islets, enhancing beta-cell hyperactivity, increasing the expression of beta-cell antigens, and resulting in pancreatic autoimmunity.<sup>196</sup> Additionally, the post-translational modifications of human islet antigens induced by local tTG increase the affinity to HLA-DQ, improving presentation to the adjacent pro-inflammatory T cells and initiating the autoimmune cascade.<sup>197</sup> Thus, gluten intake can provoke type 1 diabetes.<sup>198</sup>

Finally, applying a GFD to diabetes-prone animals reduced tTG activity in the pancreatic islets, reduced insulitis, and delayed or reduced diabetes incidence.<sup>196</sup> Recently, Hansen *et al.* substantiated these beneficial effects of gluten withdrawal in NOD mice across three generations by modulating the systemic immune system in a microbiota-independent manner, probably through epigenetic modifications.<sup>199</sup> Additionally, a GFD was shown to modulate inflammation in the salivary glands and pancreatic islets in NOD mice.<sup>200</sup>

The role of the pancreatic gluten-tTG axis in human type 1 diabetes mellitus requires further investigation.

#### **Gluten deposits in the Kidney**

The CD is associated with several renal abnormalities,<sup>201</sup> with IgA nephropathy, also known as Berger's disease, being highly affected by gluten.<sup>202</sup> It is evident that interactions between tTG and gluten peptides occur in the kidneys of IgA nephropathy patients. Indeed, tTG is overexpressed in the gut of IgA nephropathy mice, and gliadin peptides participate in the disease pathology.<sup>203</sup> The immune mechanism of gluten-induced nephropathy involves transferrin receptors, IgA1, gliadin peptides, and soluble CD89.<sup>202</sup> As proof of concept, several CD patients on GFDs showed resolution of their autoimmune kidney disease.<sup>42,204</sup> The systemic circulation of gluten peptides excreted via urine further strengthens the concept of the enteric gluten-kidney axis, where gluten withdrawal might be beneficial.

#### **Gluten deposits in the liver**

The gut-liver axis operates in CD, with several liver conditions associated with hepatic disease, ranging from isolated transaminasemia to autoimmune hepatitis.<sup>8,9,30,41,42,205,206</sup> Recently, a causal relationship was demonstrated for hepatic IgA-tTG deposits in CD patients, showing 100% sensitivity and 85% positive predictive value, establishing the association between gluten consumption,

liver IgA-tTG aggregates, and liver pathology in CD.<sup>207</sup> Not surprisingly, the resolution of liver injury and disappearance of these colocalized deposits were demonstrated with GFD in patients.<sup>207</sup>

### **Gluten deposits in the heart**

IgG and IgA immune deposition can be detected in the pericardium of individuals with gluten-dependent dermatitis herpetiformis suffering from recurrent pericarditis.<sup>208</sup> However, the gut-heart axis presents multiple clinical phenotypes in CD, including atrial fibrillation, dilated cardiomyopathy, pericarditis, myocarditis, angina pectoris, myocardial infarction, and even death from anoxic heart disease.<sup>209–211</sup> Some of these manifestations arise from hypercoagulability in CD, which is caused by multifactorial mechanisms involving nutritional deficiencies and autoantibodies.<sup>212–214</sup> Alpha-enolase, a glycolytic enzyme expressed in most tissues that plays a role in many cell functions, has been identified as an autoantigen in Hashimoto's encephalopathy. Recently, alpha-enolase has been suggested to play a role in the cardiac manifestations of CD.<sup>46</sup> Regarding cardiac involvement, CD IgA-tTG antibodies have shown strong fluorescence when applied to heart structures.<sup>215,216</sup>

### **Gluten and neurodegenerative diseases**

The well-established gut-brain axis has been thoroughly reported and reviewed.<sup>131,176,217</sup> However, the topic of gluten involvement in neurodegenerative conditions has recently generated scientific and clinical interest.<sup>8,65,66,134,217,218</sup> The current hypothesis is that indigestible luminal immunogenic gluten peptides are transported transcellularly and enter paracellularly. After crossing the blood-brain barrier, gluten peptides, cross-linked gluten complexes, gluten-induced antibodies, or gut-originated gluten-restricted CD4 T cells initiate and maintain proinflammatory cytokines, driving neurodegenerative diseases.<sup>65,66,131,134,217,218</sup> Most recently, cross-reactive antibodies between gluten and human brain epitopes have been described.<sup>44,45,219–222</sup> Specifically, cross-reactive antibodies between tTG, mTG, and amyloid-beta 1-42 have been identified,<sup>223</sup> potentially contributing to intraneuronal deposition of A-beta-P-42 in Alzheimer's disease. Similarly, cross-reactivity between tTG, mTG, wheat proteins, and alpha-synuclein has been reported,<sup>220</sup> reinforcing the role of gluten-tTG-mTG interactions in both Alzheimer's and Parkinson's diseases.

Both human endogenous and microbial exogenous transglutaminases are heavily involved in CD evolution. The tTG is the autoantigen in CD,<sup>12,13</sup> while mTG, a heavily consumed processed food additive, is described as a potential driver in CD<sup>66,94,134,224–229</sup> and systemic autoimmunity. Gluten is a prime substrate for both enzymes.<sup>66,94</sup> Gliadin-cross-linked complexes formed by these enzymes elicit high antibody levels in untreated CD children,<sup>13–16,66,96,98,99</sup> and the corresponding serological markers are very reliable for diagnosing gluten-sensitive enteropathy.<sup>14–16,97,99</sup> Finally, sequence similarity between gluten and brain epitopes was recently detected for Parkinson's disease and other neurodegenerative conditions.<sup>65,220</sup> Both mechanisms—cross-reactivity and sequence similarity between gluten peptides—contribute to molecular mimicry, which may result in neurodegenerative, neuroinflammatory, and neuropsychiatric conditions.

### **A GFD might be beneficial in many non-celiac autoimmune diseases**

If gluten is a proinflammatory and auto-immunogenic nutrient and is the offending toxic inflammatory molecule in gluten-dependent ADs, a major question arises considering its beneficial curative ef-

fects when withdrawn: Might a GFD be helpful for patients affected by non-celiac ADs? This topic was recently reviewed and summarized.<sup>41,42,55,230</sup> In a recent systematic review summarizing 83 publications, we found that 911/1,408 AD-affected patients showed improvement on a GFD. Abstaining from gluten intake was found to be efficient in 80% of the publications and clinically beneficial to 65% of the patients.<sup>42</sup> The following ADs were screened: rheumatoid arthritis, antiphospholipid syndrome, dermatomyositis, undifferentiated connective tissue disease, Raynaud's phenomenon, spondyloarthritis, psoriasis, vitiligo, pemphigus, erythema elevatum diutinum, inflammatory bowel disease, Crohn's disease, ulcerative colitis, autoimmune hepatitis, primary biliary cholangitis, primary sclerosing cholangitis, autoimmune pancreatitis, autoimmune enteropathy, thyroiditis, Graves' disease, Hashimoto's disease, type 1 diabetes, Addison's disease, autoimmune hypopituitarism, multiple sclerosis, myasthenia gravis, autoimmune myocarditis, autoimmune pericarditis, IgA nephropathy, uveitis, idiopathic thrombocytopenic purpura, and idiopathic dilated cardiomyopathy.<sup>42</sup> We concluded that a GFD might be beneficial for some patients affected by ADs. We suggested screening autoimmune patients for CD-associated antibodies, and only those who test positive should consider gluten withdrawal.<sup>42</sup> Overall, there is insufficient evidence to support a GFD for all AD patients, and official guidelines for patient selection have not yet been issued.<sup>42,55,230,231</sup> The topic is still controversial, and some studies oppose gluten withdrawal in non-celiac ADs.<sup>232–234</sup>

Logically, a GFD might counteract the harmful effects of gluten consumption. Before detailing the potential pathways and mechanisms by which gluten withdrawal might alleviate the clinical phenotype, evolution, and behavior of ADs, the following is a summary of gluten's side effects.

A challenging puzzle is the pathophysiological pathways and mechanisms by which gluten peptides induce inflammatory pathologies in various organs. Clarifying these mechanisms will improve our understanding of the beneficial effects of a GFD. The following summary (Table 1) is based on past and recent publications.<sup>6,8,9,14–16,25–30,34–39,41,42,55,65–68,70–82,89–119,121–130,134,224–230,235</sup> It should be stressed that most studies were done *ex vivo* or on animal models. Substantiation of all of them *in vivo*, on humans, is highly encouraged.

### **Potential mechanisms and pathways of GFD effectiveness in ADs**

When gluten does not enter the body, all the positive and negative effects are eliminated or avoided. Several mechanisms can prevent the inflammatory or autoimmune phenomena triggered by gut-originated gluten peptides:

1. Gluten withdrawal will eliminate or attenuate the harmful inflammatory, immunogenic, oxidative, stressogenic, dysbiotic, metabolic, and cellular consequences described in Table 1.
2. Short-chain fatty acids (SCFA) are the main microbial fermentation products in the human gut. These molecules are essential for intestinal homeostasis and the proper functioning of many protective systems in our body.<sup>76,236</sup> Untreated CD patients have disturbed microbiome diversity and composition, and their stool SCFA levels are reduced.<sup>76</sup> After one year of abstaining from gluten, their microbiome and stool SCFA content normalized.<sup>76,237</sup>
3. Tryptophan and its metabolites regulate immune functions, are essential for enteric homeostasis, and are pivotal for serotonin-dependent human behavior. Depressed and anxious CD patients have lower free tryptophan concentrations. Applying a GFD

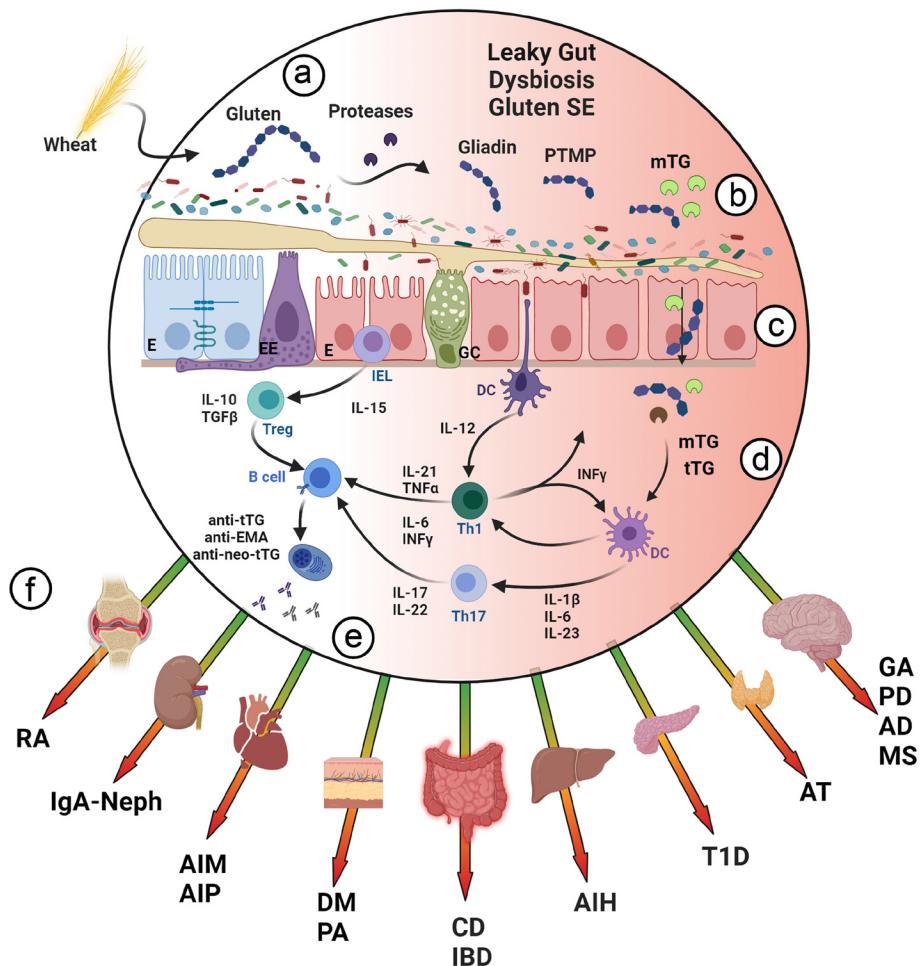
**Table 1.** Harmful effects and pathogenic mechanisms of gluten peptide-induced inflammation and cellular damage

Detrimental effects and mechanisms of gluten peptide-induced pathology	References
Pro-inflammatory	8,9,25–27,34–39,42,68,235
Drive: cytotoxicity, apoptosis, LDH; Suppress: cell viability, differentiation, RNA, DNA and glycoprotein synthesis	41,121,128–130
Cellular stress induction	42,68,70,71
Induce zonulin production	78,79,121
Pro-oxidative	42,72,121–123
Epigenetics impact	8,122,125–127
Pro-apoptotic	73,124
Impact nutrigenomics, nutrigenetics, gene expression	127
Induce dysbiosis	28–30,73–77
Increase macrophage's proinflammatory cytokine	101–103
Increase intestinal permeability	6,78–82,121
Enhance NO production	102
Immunogenic and induced antibodies	89–93
Upregulate MHCII, co-stimulatory molecules, TRLs, cytokine and chemokine production	104,105
Cross-linked to mTG immunogenicity	8,9,14–16,29,30,41,66,94,96–100
Stimulate TH1 cytokine profile	108–110
Enhance NO production	102
Increase intraepithelial lymphocyte and intestinal damage	110
Upregulate MHCII, co-stimulatory molecules, TLRs, cytokine and chemokine production	104,105
Activated intestinal CD4 <sup>+</sup> T cells, dendritic and TH17 cells, natural killer cell cytotoxicity	106,111–113
Increase expression of NKG2D and CD71	106,114
overproduction of IL-17	112,115–117
Induce TNF $\alpha$ and IL-1 $\beta$	107
Increase IL-1 $\beta$ , activate NLRP3 inflammasome	118
Enhance neutrophil migration	119

LDH- lactate dehydrogenase, MHC-major histocompatibility complex, NO-nitrous oxide, TLR- toll like receptor.

- alleviates depression and anxiety.<sup>238,239</sup> However, a recent Iranian study did not confirm this observation.<sup>240</sup> Notably, mood alterations, depression, phobia, and anxiety are prevalent in many ADs,<sup>241,242</sup> and this mechanism should be studied in non-gluten-dependent ADs.
- IgA-deficient patients are prone to ADs<sup>243</sup> and mainly to CD.<sup>244</sup> Mucosal IgA is a major immune protective mechanism, and its local production is encouraged by a high-fiber diet, local SCFA content, and a healthy microbiome. Increased production of IgA, a major mucosal and luminal immune barrier, is induced by a high-fiber diet, SCFAs, and the microbiome.<sup>245</sup> A GFD sustains a physiological microbiome, increases luminal SCFA, and combined with a high-fiber diet, could enhance enteric IgA levels.
  - GFD improves macrobiotic composition and diversity compared to untreated CD patients. The resulting higher production of SCFA, acetate, and butyrate lowers the luminal and stool pH,<sup>76,246</sup> improving colonic ecology by acting as anti-inflammatory and anti-cancer compounds.<sup>76,247,248</sup>
  - Avoiding gluten and adopting the Mediterranean diet benefit CD patients' health.<sup>62,63,249</sup> The recently adopted combination of the

- GFD-MedD pyramid avoids the harmful effects of gluten while adding the numerous benefits of the Mediterranean diet.<sup>62,250,251</sup> MedD represents a mental and physical health-protective menu that can easily be rendered gluten-free.<sup>62,63,249–251</sup> The combined diet offers higher antioxidants, anti-inflammatory nutrients, and sufficient fiber content for the CD population.<sup>64</sup>
- Gluten/gliadin, being primary substrates for tTG,<sup>94</sup> are cross-linked or deamidated by post-translational modification, losing their tolerance and becoming immune- and auto-immunogenic.<sup>29,30,100</sup> In the absence of gluten, no such reactions occur, and the body avoids gluten-dependent inflammation and tissue/organ pathologies.<sup>8,9,41,42,55,65,66</sup>
  - Cross-reactive antibodies between gluten/wheat and human tissue epitopes might induce ADs or organ pathology by molecular mimicry.<sup>44,45,220–223</sup> When no gluten peptides circulate, no cross-reactive antibodies are produced, preventing molecular mimicry.<sup>41,42,55,65,66</sup>
  - Sequence similarity between gluten and human tissue antigens has been reported recently.<sup>65,219,220</sup> The shared homology and cross-reactivity between gluten peptides and human epitopes



**Fig. 1. Pathogenic mechanisms by which gluten/gliadin peptides induce inflammation in remote organs, and the extended potential harmful effects in non-celiac ADs.** (a) Gluten is ingested and digested, reaching the gut lumen as gliadin peptides; (b) Gliadins are a prime substrate for deamidation and cross-linking by luminal transglutaminases. These post-translationally modified proteins (PTMP) increase their immunogenicity. Luminal digestive enzymes cannot further break down these protein complexes, leading to an inflammatory cascade that results in mucus disruption, dysbiosis, intestinal epithelial damage, and leaky gut; (c) Gliadin peptides and transglutaminases can potentially infiltrate through open junctions or trans-enterocytically into the lamina propria, exposing the highly immunoreactive sub-epithelium to foreign antigens or complexes; (d) In the lamina propria, dendritic cells (DCs) encounter gliadin-transglutaminase cross-linked complexes and migrate to lymph nodes as antigen-presenting cells to activate T cells. Secretion of IFN $\gamma$ , IL-17, and IL-22 by Th1 and Th17 cells activates B cells, which secrete anti-tTG, anti-neo-tTG, and anti-endomysial autoantibodies (EMA); (e) Mucosal immune cells, immunogenic modified peptides, proinflammatory cytokines, autoantibodies, and small particles that escape the immune system enter the blood vessels. They can eventually reach remote organs and trigger an autoimmune response; (f) Some examples of inflammatory conditions that can be affected by the presence of gliadin peptides and transglutaminases or cross-linked complexes in peripheral organs. AD, Alzheimer's Disease; AIH, Autoimmune Hepatitis; AIM, Autoimmune Myocarditis; AIP, Autoimmune Pericarditis; AT, Autoimmune Thyroiditis; CD, Celiac disease; DM, Dermatomyositis; GA, Gluten Ataxia; IBD, Inflammatory Bowel Diseases; IEL, intraepithelial lymphocytes.; IgA-Neph, IgA nephropathy (Berger's disease); MS, Multiple Sclerosis; mTG, microbial transglutaminase; PA, Psoriatic Arthritis; PD, Parkinson's Disease; RA, Rheumatoid Arthritis; T1D, Type 1 Diabetes; tTG, tissue transglutaminase.

reinforce the molecular mimicry pathway toward inflammation and end-organ dysfunction. A GFD prevents these phenomena by curtailing shared sequences and cross-reactivity. Intriguingly, cross-reactivity and sequence similarity have recently been reported between various human antigens and a family member of tTG, namely, microbial transglutaminase.<sup>252</sup>

10. Leaky gut is reported in many, *in vivo* and *ex vivo*, metabolic, inflammatory, and ADs.<sup>235,253–255</sup> Gluten is a major disruptor of enteric tight junction functional integrity.<sup>41,42,65,66,235,254,255</sup> Gluten avoidance might protect the body from this abnormality. However, the enigma of “Gluten: yes, no, maybe” is far from being resolved.<sup>256</sup>

The pathogenic mechanisms by which gluten/gliadin peptides induce inflammation in remote organs, and the extended potential harmful effects in non-celiac ADs are described in Figure 1

## Conclusion

Gluten has many side effects that compromise human health, not only in gluten-dependent conditions but also in non-gluten-related chronic diseases. After entering the gut lumen, undigestible gluten peptides are modified by luminal mTG or transported through the enteric epithelium to meet mucosal immune cells or distributed systemically to remote organs where they encounter tTG. The

modified peptides become immunogenic and pro-inflammatory, inducing organ dysfunction and pathology. A GFD can prevent these phenomena by multiple mechanisms: suppressing gluten-associated detrimental effects, improving the microbiome/dysbiome ratio, avoiding post-translational modification of gluten peptides, preventing cross-reactivity and sequence similarity between gluten and human epitopes, and reducing gut leakage.

As proof of concept, gluten withdrawal alleviates disease activity in multiple chronic inflammatory, metabolic, autoimmune conditions, and even neurodegeneration. However, caution is needed. GFD consumers should be aware of the disadvantages of a gluten-restricted diet. It is advised to combine a GFD with the Mediterranean diet to harness the advantages of both. Before recommending a GFD for non-gluten-dependent conditions, patients should be assessed for gut symptomatology and screened for celiac-associated antibodies. Notably, this topic is still under discussion and is not included in the guidelines of professional decision-making societies.

It is hoped that this narrative review will encourage the scientific, nutritional, and medical communities to further explore the mechanisms by which gluten peptides induce inflammation and end-organ damage. Understanding these pathways will clarify gluten's role in the induction of human chronic inflammatory diseases.

## Acknowledgments

None.

## Funding

None.

## Conflict of interest

AV is the CEO of Immunosciences Lab., Inc. The authors have no other conflict of interests to declare.

## Author contributions

Screening the literature (AL, CB), designing and writing the manuscript (AL), editing and revising the manuscript, designing the figure with BioRender.com permission (CB), writing part of the manuscript, revising the manuscript, and summarizing the cross-reactivity results (AV). The three authors agreed to the published version of the manuscript.

## References

- [1] Furman D, Campisi J, Verdin E, Carrera-Bastos P, Targ S, Franceschi C, et al. Chronic inflammation in the etiology of disease across the life span. *Nat Med* 2019;25(12):1822–1832. doi:10.1038/s41591-019-0675-0, PMID:31806905.
- [2] Sears B. Anti-inflammatory Diets. *J Am Coll Nutr* 2015;34(Suppl 1):14–21. doi:10.1080/07315724.2015.1080105, PMID:26400429.
- [3] Haß U, Herpich C, Norman K. Anti-Inflammatory Diets and Fatigue. *Nutrients* 2019;11(10):2315. doi:10.3390/nu11102315, PMID:31574939.
- [4] Gill PA, Inniss S, Kumagai T, Rahman FZ, Smith AM. The Role of Diet and Gut Microbiota in Regulating Gastrointestinal and Inflammatory Disease. *Front Immunol* 2022;13:866059. doi:10.3389/fimmu.2022.866059, PMID:35450067.
- [5] Malesza IJ, Malesza M, Walkowiak J, Mussin N, Walkowiak D, Aringazina R, et al. High-Fat, Western-Style Diet, Systemic Inflammation, and Gut Microbiota: A Narrative Review. *Cells* 2021;10(11):3164. doi:10.3390/cells10113164, PMID:34831387.
- [6] Lerner A, Matthias T. Changes in intestinal tight junction permeability associated with industrial food additives explain the rising incidence of autoimmune disease. *Autoimmun Rev* 2015;14(6):479–489. doi:10.1016/j.autrev.2015.01.009, PMID:25676324.
- [7] Liu W, Chen X, Li H, Zhang J, An J, Liu X. Anti-Inflammatory Function of Plant-Derived Bioactive Peptides: A Review. *Foods* 2022;11(15):2361. doi:10.3390/foods11152361, PMID:35954128.
- [8] Aaron L, Torsten M, Patricia W. Autoimmunity in celiac disease: Extra-intestinal manifestations. *Autoimmun Rev* 2019;18(3):241–246. doi:10.1016/j.autrev.2018.09.010, PMID:30639642.
- [9] Lerner A, Matthias T. GUT-the Trojan Horse in remote organs' Autoimmunity. *J Clin Cell Immunol* 2016;7:1–10. doi:10.4172/2155-9899.1000401.
- [10] Catassi C, Verdu EF, Bai JC, Lionetti E. Celiac disease. *Lancet* 2022;399(10344):2413–2426. doi:10.1016/S0140-6736(22)00794-2, PMID:35691302.
- [11] Lerner A. The Enigma of Oats in Nutritional Therapy for Celiac Disease. *Int J Celiac Dis* 2014;2:110–114. doi:10.12691/IJCD-2-3-1.
- [12] Reif S, Lerner A. Tissue transglutaminase—the key player in celiac disease: a review. *Autoimmun Rev* 2004;3(1):40–45. doi:10.1016/S1568-9972(03)00065-X, PMID:14871648.
- [13] Lerner A, Neidhöfer S, Matthias T. Transglutaminase 2 and anti transglutaminase 2 autoantibodies in celiac disease and beyond: TG2 double-edged sword: Gut and extraintestinal involvement. *Immunome Res* 2015;11:1–4. doi:10.4172/1745-7580.10000101.
- [14] Lerner A, Ramesh A, Matthias T. Serologic Diagnosis of Celiac Disease: New Biomarkers. *Gastroenterol Clin North Am* 2019;48(2):307–317. doi:10.1016/j.gtc.2019.02.009, PMID:31046977.
- [15] Lerner A, Jeremias P, Neidhofer S, Matthias T. Comparison of the Reliability of 17 Celiac Disease Associated Bio-Markers to Reflect Intestinal Damage. *J Clin Cell Immunol* 2017;8:486. doi:10.4172/2155-9899.1000486.
- [16] Agardh D, Matthias T, Wusterhausen P, Neidhöfer S, Heller A, Lerner A. Antibodies against neo-epitope of microbial and human transglutaminase complexes as biomarkers of childhood celiac disease. *Clin Exp Immunol* 2020;199(3):294–302. doi:10.1111/cei.13394, PMID:31663117.
- [17] Lerner A, Blank M, Shoenfeld Y. Celiac disease and autoimmunity. *Isr J Med Sci* 1996;32:33–36.
- [18] Sollid LM, Jabri B. Is celiac disease an autoimmune disorder? *Curr Opin Immunol* 2005;17(6):595–600. doi:10.1016/j.co.2005.09.015, PMID:16214317.
- [19] Troncone R, Discepolo V. Celiac disease and autoimmunity. *J Pediatr Gastroenterol Nutr* 2014;59(Suppl 1):S9–S11. doi:10.1097/01.mpg.0000450394.30780.ea, PMID:24979198.
- [20] Villanacci V, Vanoli A, Leoncini G, Arpa G, Salvato T, Bonetti LR, et al. Celiac disease: histology-differential diagnosis-complications. A practical approach. *Pathologica* 2020;112(3):186–196. doi:10.32074/1591-951X-157, PMID:33179621.
- [21] Lerner A, Matthias T. Intraepithelial lymphocyte normal cut-off level in celiac disease: The debate continues. *Int J Celiac Dis* 2016;4:4–6. doi:10.12691/ijcd-4-1-1.
- [22] Lindeman I, Sollid LM. Single-cell approaches to dissect adaptive immune responses involved in autoimmunity: the case of celiac disease. *Mucosal Immunol* 2022;15(1):51–63. doi:10.1038/s41385-021-00452-0, PMID:34531547.
- [23] Kim SM, Mayassi T, Jabri B. Innate immunity: actuating the gears of celiac disease pathogenesis. *Best Pract Res Clin Gastroenterol* 2015;29(3):425–435. doi:10.1016/j.bpg.2015.05.001, PMID:26060107.
- [24] Anderson RP. Innate and adaptive immunity in celiac disease. *Curr Opin Gastroenterol* 2020;36(6):470–478. doi:10.1097/MOG.0000000000000672, PMID:32889822.
- [25] Voisine J, Abadie V. Interplay Between Gluten, HLA, Innate and Adaptive Immunity Orchestrates the Development of Coeliac Disease. *Front Immunol* 2021;12:674313. doi:10.3389/fimmu.2021.674313, PMID:34149709.
- [26] Vincentini O, Maialetti F, Gonnelli E, Silano M. Gliadin-dependent cytokine production in a bidimensional cellular model of celiac intestinal mucosa. *Clin Exp Med* 2015;15(4):447–454. doi:10.1007/s10238-014-0325-2, PMID:25447073.

- [27] Garrote JA, Gómez-González E, Bernardo D, Arranz E, Chirdo F. Celiac disease pathogenesis: the proinflammatory cytokine network. *J Pediatr Gastroenterol Nutr* 2008;47(Suppl 1):S27–S32. doi:10.1097 MPG.0b013e31818fb9, PMID:18667914.
- [28] Wu X, Qian L, Liu K, Wu J, Shan Z. Gastrointestinal microbiome and gluten in celiac disease. *Ann Med* 2021;53(1):1797–1805. doi:10.1080/07853890.2021.1990392, PMID:34647492.
- [29] Lerner A, Aminov R, Matthias T. Dysbiosis May Trigger Autoimmune Diseases via Inappropriate Post-Translational Modification of Host Proteins. *Front Microbiol* 2016;7:84. doi:10.3389/fmicb.2016.00084, PMID:26903965.
- [30] Lerner A, Aminov R, Matthias T. Transglutaminases in Dysbiosis As Potential Environmental Drivers of Autoimmunity. *Front Microbiol* 2017;8:66. doi:10.3389/fmicb.2017.00066, PMID:28174571.
- [31] Lerner A, Matthias T. The Yin and Yang of dietary gluten transgressions in real-life scenarios of celiac patients. *BMC Med* 2020;18(1):70. doi:10.1186/s12916-020-01535-8, PMID:32156283.
- [32] Samasca G, Lerner A, Girbovan A, Sur G, Lupan I, Makovicky P, et al. Challenges in gluten-free diet in coeliac disease: Prague consensus. *Eur J Clin Invest* 2017;47(5):394–397. doi:10.1111/eci.12755, PMID:28369858.
- [33] Huj Joel IA, Murray JA. Refractory Celiac Disease. *Curr Gastroenterol Rep* 2020;22(4):18. doi:10.1007/s11894-020-0756-8, PMID:32185560.
- [34] Patel N, Robert ME. Frontiers in Celiac Disease: Where Autoimmunity and Environment Meet. *Am J Surg Pathol* 2022;46(1):e43–e54. doi:10.1097/PAS.0000000000001639, PMID:33739793.
- [35] Mayassi T, Ladell K, Gudjonson H, McLaren JE, Shaw DG, Tran MT, et al. Chronic Inflammation Permanently Reshapes Tissue-Resident Immunity in Celiac Disease. *Cell* 2019;176(5):967–981.e19. doi:10.1016/j.cell.2018.12.039, PMID:30739797.
- [36] Auricchio R, Calabrese I, Galatola M, Cielo D, Carbone F, Mancuso M, et al. Gluten consumption and inflammation affect the development of celiac disease in at-risk children. *Sci Rep* 2022;12(1):5396. doi:10.1038/s41598-022-09232-7, PMID:35354862.
- [37] Levescot A, Malamut G, Cerf-Bensussan N. Immunopathogenesis and environmental triggers in coeliac disease. *Gut* 2022;71(11):2337–2349. doi:10.1136/gutjnl-2021-326257, PMID:35879049.
- [38] Porpora M, Conte M, Lania G, Bellomo C, Rapacciuolo L, Chirdo FG, et al. Inflammation Is Present, Persistent and More Sensitive to Pro-inflammatory Triggers in Celiac Disease Enterocytes. *Int J Mol Sci* 2022;23(4):1973. doi:10.3390/ijms23041973, PMID:35216089.
- [39] de Punder K, Pruijboom L. The dietary intake of wheat and other cereal grains and their role in inflammation. *Nutrients* 2013;5(3):771–787. doi:10.3390/nu5030771, PMID:23482055.
- [40] Zingone F. Grain Intake and Human Health. *Nutrients* 2020;12(12):3733. doi:10.3390/nu12123733, PMID:33291526.
- [41] Lerner A, Shoenfeld Y, Matthias T. Adverse effects of gluten ingestion and advantages of gluten withdrawal in nonceliac autoimmune disease. *Nutr Rev* 2017;75(12):1046–1058. doi:10.1093/nutrit/nux054, PMID:29202198.
- [42] Lerner A, Freire de Carvalho J, Kotrova A, Shoenfeld Y. Gluten-free diet can ameliorate the symptoms of non-celiac autoimmune diseases. *Nutr Rev* 2022;80(3):525–543. doi:10.1093/nutrit/nuab039, PMID:34338776.
- [43] Garutti M, Nevola G, Mazzeo R, Cucciniello L, Totaro F, Bertuzzi CA, et al. The Impact of Cereal Grain Composition on the Health and Disease Outcomes. *Front Nutr* 2022;9:888974. doi:10.3389/fnut.2022.888974, PMID:35711559.
- [44] Vojdani A. Reaction of food-specific antibodies with different tissue antigens. *Int J Food Sci Technol* 2020;55:1800–1815. doi:10.1111/ijfs.14467.
- [45] Vojdani A, Gushgari LR, Vojdani E. Interaction between food antigens and the immune system: Association with autoimmune disorders. *Autoimmun Rev* 2020;19(3):102459. doi:10.1016/j.autrev.2020.102459, PMID:31917265.
- [46] Lerner A, Sobolevskaia P, Churilov L, Shoenfeld Y. Alpha-enolase involvement in intestinal and extraintestinal manifestations of celiac disease. *J Transl Autoimmun* 2021;4:100109. doi:10.1016/j.jtau.2021.100109, PMID:34189450.
- [47] FAOSTAT Food and Agriculture Organization of the United Nations. Available from: <http://www.fao.org/faostat/en/#data>. Accessed October 13, 2022.
- [48] <https://www.globenewswire.com/news-release/2020/06/30/2055191/0/en/Global-Vital-Wheat-Gluten-Industry-Assessment-2018-2020-2024-2027.html>.
- [49] Harari YN. *Sapiens: A Brief History of Humankind*. Oxford: Signal; 2017.
- [50] Wieser H, Koehler P, Scherf KA. The Two Faces of Wheat. *Front Nutr* 2020;7:517313. doi:10.3389/fnut.2020.517313, PMID:33195360.
- [51] Vojdani A, Tarash I. Cross-Reaction between Gliadin and Different Food and Tissue Antigens. *Food Nutr Sci* 2013;04:20–32. doi:10.4236/fns.2013.41005.
- [52] Shewry P. What Is Gluten-Why Is It Special? *Front Nutr* 2019;6:101. doi:10.3389/fnut.2019.000101, PMID:31334243.
- [53] Lerner A, O'Bryan T, Matthias T. Navigating the Gluten-Free Boom: The Dark Side of Gluten Free Diet. *Front Pediatr* 2019;7:414. doi:10.3389/fped.2019.00414, PMID:31681712.
- [54] Shmerling RH. Harvard Health Publishing. Harvard Medical School. Ditch the Gluten, Improve Your Health? April 14, 2022. <https://www.health.harvard.edu/staying-healthy/ditch-the-gluten-improve-your-health> (accessed October 13, 2022).
- [55] Lerner A, Ramesh A, Matthias T. Going gluten free in non-celiac autoimmune diseases: the missing ingredient. *Expert Rev Clin Immunol* 2018;14(11):873–875. doi:10.1080/1744666X.2018.1524757, PMID:30220227.
- [56] Lerner A. New therapeutic strategies for celiac disease. *Autoimmun Rev* 2010;9(3):144–147. doi:10.1016/j.autrev.2009.05.002, PMID:19427921.
- [57] Lerner A, Matthias T. Gluten-free diet tough alley in torrid time. *Int J Celiac Dis* 2017;5:50–5. doi:10.12691/ijcd-5-2-4.
- [58] Sergi C, Villanacci V, Carroccio A. Non-celiac wheat sensitivity: rationality and irrationality of a gluten-free diet in individuals affected with non-celiac disease: a review. *BMC Gastroenterol* 2021;21(1):5. doi:10.1186/s12876-020-01568-6, PMID:33407153.
- [59] Siddiqui UN, Pervaiz A, Khan ZB, Sultana T. Diagnostic Dilemma, Possible Non-celiac Gluten Sensitivity: Consideration in Approach and Management. *Cureus* 2022;14(5):e25302. doi:10.7759/cureus.25302, PMID:35774680.
- [60] Reese I, Schäfer C, Kleine-Tebbe J, Ahrens B, Bachmann O, Ballmer-Weber B, et al. Non-celiac gluten/wheat sensitivity (NCGS)-a currently undefined disorder without validated diagnostic criteria and of unknown prevalence: Position statement of the task force on food allergy of the German Society of Allergology and Clinical Immunology (DGAKI). *Allergo J Int* 2018;27(5):147–151. doi:10.1007/s40629-018-0070-2, PMID:30294520.
- [61] Mumolo MG, Rettura F, Melissari S, Costa F, Ricchiuti A, Ceccarelli L, et al. Is Gluten the Only Culprit for Non-Celiac Gluten/Wheat Sensitivity? *Nutrients* 2020;12(12):3785. doi:10.3390/nu12123785, PMID:33321805.
- [62] Bascuñán KA, Elli L, Vecchi M, Scricciolo A, Mascaretti F, Parisi M, et al. Mediterranean Gluten-Free Diet: Is It a Fair Bet for the Treatment of Gluten-Related Disorders? *Front Nutr* 2020;7:583981. doi:10.3389/fnut.2020.583981, PMID:33344491.
- [63] Nestares T, Martín-Masot R, de Teresa C, Bonillo R, Maldonado J, Flor-Alemany M, et al. Influence of Mediterranean Diet Adherence and Physical Activity on Bone Health in Celiac Children on a Gluten-Free Diet. *Nutrients* 2021;13(5):1636. doi:10.3390/nu13051636, PMID:34068001.
- [64] Cenni S, Sesenna V, Boiardi G, Casertano M, Di Nardo G, Esposito S, et al. The Mediterranean Diet in Paediatric Gastrointestinal Disorders. *Nutrients* 2022;15(1):79. doi:10.3390/nu15010079, PMID:36615737.
- [65] Lerner A, Benzi C. “Let Food Be Thy Medicine”: Gluten and Potential Role in Neurodegeneration. *Cells* 2021;10(4):756. doi:10.3390/cells10040756, PMID:33808124.
- [66] Lerner A, Matthias T. Gluten and Autoimmunogenesis. In *Mosaic of Autoimmunity: The Novel Factors of Autoimmune Diseases Revisited*. 2nd ed. Academic Press; 2019. doi:10.1016/B978-0-12-814307-0.00032-3.
- [67] Lammers KM, Herrera MG, Dodero VI. Translational Chemistry Meets Gluten-Related Disorders. *ChemistryOpen* 2018;7(3):217–232. doi:10.1002/open.201700197, PMID:29531885.

- [68] Chirdo FG, Auricchio S, Troncone R, Barone MV. The gliadin p31-43 peptide: Inducer of multiple proinflammatory effects. *Int Rev Cell Mol Biol* 2021;358:165–205. doi:10.1016/bs.ircmb.2020.10.003, PMID:33707054.
- [69] Barone MV, Auricchio R, Nanayakkara M, Greco L, Troncone R, Auricchio S. Pivotal Role of Inflammation in Celiac Disease. *Int J Mol Sci* 2022;23(13):7177. doi:10.3390/ijms23137177, PMID:35806180.
- [70] Goel G, Tye-Din JA, Qiao SW, Russell AK, Mayassi T, Ciszewski C, et al. Cytokine release and gastrointestinal symptoms after gluten challenge in celiac disease. *Sci Adv* 2019;5(8):eaaw7756. doi:10.1126/sciadv.aaw7756, PMID:31457091.
- [71] Conte M, Nigro F, Porpora M, Bellomo C, Furone F, Budelli AL, et al. Gliadin Peptide P31-43 Induces mTOR/NFκB Activation and Reduces Autophagy: The Role of Lactobacillus paracasei CBA L74 Postbiotic. *Int J Mol Sci* 2022;23(7):3655. doi:10.3390/ijms23073655, PMID:35409015.
- [72] Monguzzi E, Marabini L, Elli L, Vaira V, Ferrero S, Ferretti F, et al. Gliadin effect on the oxidative balance and DNA damage: An in-vitro, ex-vivo study. *Dig Liver Dis* 2019;51(1):47–54. doi:10.1016/j.dld.2018.06.020, PMID:30055963.
- [73] Perez F, Ruera CN, Miculan E, Carasi P, Chirdo FG. Programmed Cell Death in the Small Intestine: Implications for the Pathogenesis of Celiac Disease. *Int J Mol Sci* 2021;22(14):7426. doi:10.3390/ijms22147426, PMID:34299046.
- [74] Chander AM, Yadav H, Jain S, Bhadada SK, Dhawan DK. Cross-Talk Between Gluten, Intestinal Microbiota and Intestinal Mucosa in Celiac Disease: Recent Advances and Basis of Autoimmunity. *Front Microbiol* 2018;9:2597. doi:10.3389/fmicb.2018.02597, PMID:30443241.
- [75] Ren Z, Pan LL, Huang Y, Chen H, Liu Y, Liu H, et al. Gut microbiota-CRAMP axis shapes intestinal barrier function and immune responses in dietary gluten-induced enteropathy. *EMBO Mol Med* 2021;13(8):e14059. doi:10.15252/emmm.202114059, PMID:34125490.
- [76] Lerner A, Patricia J, Matthias T. Nutrients, Bugs and Us: The Short-chain Fatty Acids Story in Celiac Disease. *Int J Celiac Dis* 2016;4:92–4. doi:10.12691/IJCD-4-3-12.
- [77] Lerner A, Arleevskaya M, Schmiedl A, Matthias T. Microbes and Viruses Are Bugging the Gut in Celiac Disease. Are They Friends or Foes? *Front Microbiol* 2017;8:1392. doi:10.3389/fmicb.2017.01392, PMID:28824555.
- [78] Guerreiro CS, Calado Â, Sousa J, Fonseca JE. Diet, Microbiota, and Gut Permeability-The Unknown Triad in Rheumatoid Arthritis. *Front Med (Lausanne)* 2018;5:349. doi:10.3389/fmed.2018.00349, PMID:30619860.
- [79] Wood Heickman LK, DeBoer MD, Fasano A. Zonulin as a potential putative biomarker of risk for shared type 1 diabetes and celiac disease autoimmunity. *Diabetes Metab Res Rev* 2020;36(5):e3309. doi:10.1002/dmrr.3309, PMID:32162764.
- [80] An J, Liu Y, Wang Y, Fan R, Hu X, Zhang F, et al. The Role of Intestinal Mucosal Barrier in Autoimmune Disease: A Potential Target. *Front Immunol* 2022;13:871713. doi:10.3389/fimmu.2022.871713, PMID:35844539.
- [81] Jauregi-Miguel A. The tight junction and the epithelial barrier in coeliac disease. *Int Rev Cell Mol Biol* 2021;358:105–132. doi:10.1016/bs.ircmb.2020.09.010, PMID:33707052.
- [82] Drago S, El Asmar R, Di Pierro M, Grazia Clemente M, Tripathi A, Sapone A, et al. Gliadin, zonulin and gut permeability: Effects on celiac and non-celiac intestinal mucosa and intestinal cell lines. *Scand J Gastroenterol* 2006;41(4):408–419. doi:10.1080/00365520500235334, PMID:16635908.
- [83] Hoilat GJ, Altowairqi AK, Ayas MF, Alhaddab NT, Alnujaidi RA, Alharbi HA, et al. Larazotide acetate for treatment of celiac disease: A systematic review and meta-analysis of randomized controlled trials. *Clin Res Hepatol Gastroenterol* 2022;46(1):101782. doi:10.1016/j.clinre.2021.101782, PMID:34339872.
- [84] Matei DE, Menon M, Alber DG, Smith AM, Nedjat-Shokouhi B, Fasano A, et al. Intestinal barrier dysfunction plays an integral role in arthritis pathology and can be targeted to ameliorate disease. *Med* 2021;2(7):864–883.e9. doi:10.1016/j.medj.2021.04.013, PMID:34296202.
- [85] Yonker LM, Gilboa T, Ogata AF, Senussi Y, Lazarovits R, Boribong BP, et al. Multisystem inflammatory syndrome in children is driven by zonulin-dependent loss of gut mucosal barrier. *J Clin Invest* 2021;131(14):149633. doi:10.1172/JCI149633, PMID:34032635.
- [86] Slifer ZM, Krishnan BR, Madan J, Blikslager AT. Larazotide acetate: a pharmacological peptide approach to tight junction regulation. *Am J Physiol Gastrointest Liver Physiol* 2021;320(6):G983–G989. doi:10.1152/ajpgi.00386.2020, PMID:33881350.
- [87] Di Micco S, Musella S, Sala M, Scala MC, Andrei G, Snoeck R, et al. Peptide Derivatives of the Zonulin Inhibitor Larazotide (AT1001) as Potential Anti SARS-CoV-2: Molecular Modelling, Synthesis and Bioactivity Evaluation. *Int J Mol Sci* 2021;22(17):9427. doi:10.3390/ijms22179427, PMID:34502335.
- [88] Ailioaie LM, Ailioaie C, Litscher G, Chiran DA. Celiac Disease and Targeting the Molecular Mechanisms of Autoimmunity in COVID Pandemic. *Int J Mol Sci* 2022;23(14):7719. doi:10.3390/ijms23147719, PMID:35887067.
- [89] Gillett HR, Freeman HJ. Serological testing in screening for adult celiac disease. *Can J Gastroenterol* 1999;13(3):265–269. doi:10.1155/1999/194367, PMID:10331939.
- [90] Tucker NT, Barghuthy FS, Prihoda TJ, Kumar V, Lerner A, Lebenthal E. Antigliadin antibodies detected by enzyme-linked immunosorbent assay as a marker of childhood celiac disease. *J Pediatr* 1988;113(2):286–289. doi:10.1016/s0022-3476(88)80266-x, PMID:3397791.
- [91] Lerner A, Kumar V, Iancu TC. Immunological diagnosis of childhood coeliac disease: comparison between antigliadin, antireticulin and antiendomysium antibodies. *Clin Exp Immunol* 1994;95(1):78–82. doi:10.1111/j.1365-2249.1994.tb06018.x, PMID:8287612.
- [92] Lerner A, Lebenthal E. The controversy of the use of anti-gluten antibody (AGA) as a diagnostic tool in celiac disease. *J Pediatr Gastroenterol Nutr* 1991;12(4):407–409. doi:10.1097/00005176-199105000-00001, PMID:1865276.
- [93] Vojdani A. Detection of IgE, IgG, IgA and IgM antibodies against raw and processed food antigens. *Nutr Metab (Lond)* 2009;6:22. doi:10.1186/1743-7075-6-22, PMID:19435515.
- [94] Lerner A, Matthias T. Possible association between celiac disease and bacterial transglutaminase in food processing: a hypothesis. *Nutr Rev* 2015;73(8):544–552. doi:10.1093/nutrit/nuv011, PMID:26084478.
- [95] Matthias T, Jeremias P, Neidhöfer S, Lerner A. The industrial food additive, microbial transglutaminase, mimics tissue transglutaminase and is immunogenic in celiac disease patients. *Autoimmun Rev* 2016;15(12):1111–1119. doi:10.1016/j.autrev.2016.09.011, PMID:27640315.
- [96] Lerner A. More novel diagnostic antibodies for celiac disease. *Expert Rev Gastroenterol Hepatol* 2016;10(7):767–768. doi:10.1080/17474124.2016.1175300, PMID:27237317.
- [97] Lerner A, Matthias T. Food Industrial Microbial Transglutaminase in Celiac Disease: Treat or Trick. *Int J Celiac Dis* 2015;3:1–6. doi:10.12691/IJCD-3-1-10.
- [98] Matthias T, Neidhöfer S, Pfeiffer S, Prager K, Reuter S, Gershwin ME. Novel trends in celiac disease. *Cell Mol Immunol* 2011;8(2):121–125. doi:10.1038/cmi.2010.68, PMID:21278765.
- [99] Lerner A, Jeremias P, Neidhöfer S, Matthias T. Antibodies against neo-epitope tTG complexed to gliadin are different and more reliable than anti-tTG for the diagnosis of pediatric celiac disease. *J Immunol Methods* 2016;429:15–20. doi:10.1016/j.jim.2015.12.006, PMID:26684936.
- [100] Lerner A, Matthias T. Rheumatoid arthritis-celiac disease relationship: joints get that gut feeling. *Autoimmun Rev* 2015;14(11):1038–1047. doi:10.1016/j.autrev.2015.07.007, PMID:26190704.
- [101] Thomas KE, Sapone A, Fasano A, Vogel SN. Gliadin stimulation of murine macrophage inflammatory gene expression and intestinal permeability are MyD88-dependent: role of the innate immune response in Celiac disease. *J Immunol* 2006;176(4):2512–2521. doi:10.4049/jimmunol.176.4.2512, PMID:16456012.
- [102] Tucková L, Flegelová Z, Tlaskalová-Hogenová H, Zídek Z. Activation of macrophages by food antigens: enhancing effect of gluten on nitric oxide and cytokine production. *J Leukoc Biol* 2000;67(3):312–318. doi:10.1002/jlb.67.3.312, PMID:10733090.
- [103] Tucková L, Novotná J, Novák P, Flegelová Z, Kveton T, Jelíková L, et al. Activation of macrophages by gliadin fragments: isolation and characterization of active peptide. *J Leukoc Biol* 2002;71(4):625–631. PMID:11927649.

- [104] Palová-Jelínková L, Rozková D, Pecharová B, Bártová J, Sedivá A, Tlaskalová-Hogenová H, et al. Gliadin fragments induce phenotypic and functional maturation of human dendritic cells. *J Immunol* 2005;175(10):7038–7045. doi:10.4049/jimmunol.175.10.7038, PMID:16272365.
- [105] Ciccocioppo R, Rossi M, Pesce I, Ricci G, Millimaggi D, Maurano F, et al. Effects of gliadin stimulation on bone marrow-derived dendritic cells from HLA-DQ8 transgenic MICE. *Dig Liver Dis* 2008;40(12):927–935. doi:10.1016/j.dld.2008.05.005, PMID:18567549.
- [106] Larsen J, Dall M, Antvorskov JC, Weile C, Engkilde K, Josefson K, et al. Dietary gluten increases natural killer cell cytotoxicity and cytokine secretion. *Eur J Immunol* 2014;44(10):3056–3067. doi:10.1002/eji.201344264, PMID:25043259.
- [107] Gujral N, Suh JW, Sunwoo HH. Effect of anti-gliadin IgY antibody on epithelial intestinal integrity and inflammatory response induced by gliadin. *BMC Immunol* 2015;16:41. doi:10.1186/s12865-015-0104-1, PMID:26156219.
- [108] Scott FW, Rowsell P, Wang GS, Burghardt K, Kolb H, Flohé S. Oral exposure to diabetes-promoting food or immunomodulators in neonates alters gut cytokines and diabetes. *Diabetes* 2002;51(1):73–78. doi:10.2337/diabetes.51.1.73, PMID:11756325.
- [109] Scott FW, Cloutier HE, Kleemann R, Wöhrer-Pagenstert U, Rowsell P, Modler HW, et al. Potential mechanisms by which certain foods promote or inhibit the development of spontaneous diabetes in BB rats: dose, timing, early effect on islet area, and switch in infiltrate from Th1 to Th2 cells. *Diabetes* 1997;46(4):589–598. doi:10.2337/diab.46.4.589, PMID:9075798.
- [110] Stěpánková R, Tlaskalová-Hogenová H, Sinkora J, Jodl J, Fric P. Changes in jejunal mucosa after long-term feeding of germfree rats with gluten. *Scand J Gastroenterol* 1996;31(6):551–557. doi:10.3109/00365529609009127, PMID:8789893.
- [111] Flohé SB, Wasmuth HE, Kerad JB, Beales PE, Pozzilli P, Elliott RB, et al. A wheat-based, diabetes-promoting diet induces a Th1-type cytokine bias in the gut of NOD mice. *Cytokine* 2003;21(3):149–154. doi:10.1016/s1043-4666(02)00486-6, PMID:12697153.
- [112] Alam C, Valkonen S, Palagani V, Jalava J, Eerola E, Hänninen A. Inflammatory tendencies and overproduction of IL-17 in the colon of young NOD mice are counteracted with diet change. *Diabetes* 2010;59(9):2237–2246. doi:10.2337/db10-0147, PMID:20547977.
- [113] Larsen J, Weile C, Antvorskov JC, Engkilde K, Nielsen SM, Josefson K, et al. Effect of dietary gluten on dendritic cells and innate immune subsets in BALB/c and NOD mice. *PLoS One* 2015;10(3):e0118618. doi:10.1371/journal.pone.0118618, PMID:25738288.
- [114] Adlercreutz EH, Weile C, Larsen J, Engkilde K, Agardh D, Buschard K, et al. A gluten-free diet lowers NKG2D and ligand expression in BALB/c and non-obese diabetic (NOD) mice. *Clin Exp Immunol* 2014;177(2):391–403. doi:10.1111/cei.12340, PMID:24673402.
- [115] Antvorskov JC, Fundova P, Buschard K, Funda DP. Impact of dietary gluten on regulatory T cells and Th17 cells in BALB/c mice. *PLoS One* 2012;7(3):e33315. doi:10.1371/journal.pone.0033315, PMID:22428018.
- [116] Antvorskov JC, Fundova P, Buschard K, Funda DP. Dietary gluten alters the balance of pro-inflammatory and anti-inflammatory cytokines in T cells of BALB/c mice. *Immunology* 2013;138(1):23–33. doi:10.1111/imm.12007, PMID:22913724.
- [117] Bernardo D, Garrote JA, Fernández-Salazar L, Riestra S, Arranz E. Is gliadin really safe for non-coeliac individuals? Production of interleukin 15 in biopsy culture from non-coeliac individuals challenged with gliadin peptides. *Gut* 2007;56(6):889–890. doi:10.1136/gut.2006.118265, PMID:17519496.
- [118] Palová-Jelínková L, Dáňová K, Drašarová H, Dvořák M, Funda DP, Fundová P, et al. Pepsin digest of wheat gliadin fraction increases production of IL-1β via TLR4/MyD88/TRIF/MAPK/NF-κB signaling pathway and an NLRP3 inflammasome activation. *PLoS One* 2013;8(4):e62426. doi:10.1371/journal.pone.0062426, PMID:23658628.
- [119] Lammers KM, Chieppa M, Liu L, Liu S, Omatsu T, Janka-Junttila M, et al. Gliadin Induces Neutrophil Migration via Engagement of the Formyl Peptide Receptor, FPR1. *PLoS One* 2015;10(9):e0138338. doi:10.1371/journal.pone.0138338, PMID:26378785.
- [120] Hudson DA, Cornell HJ, Purdham DR, Rolles CJ. Non-specific cytotoxicity of wheat gliadin components towards cultured human cells. *Lancet* 1976;1(7955):339–341. doi:10.1016/s0140-6736(76)90089-1, PMID:54743.
- [121] Dolfini E, Elli L, Roncoroni L, Costa B, Colleoni MP, Lorusso V, et al. Damaging effects of gliadin on three-dimensional cell culture model. *World J Gastroenterol* 2005;11(38):5973–5977. doi:10.3748/wjg.v11.i38.5973, PMID:16273608.
- [122] Trivedi MS, Shah JS, Al-Mughairy S, Hodgson NW, Simms B, Trooskens GA, et al. Food-derived opioid peptides inhibit cysteine uptake with redox and epigenetic consequences. *J Nutr Biochem* 2014;25(10):1011–1018. doi:10.1016/j.jnubio.2014.05.004, PMID:25018147.
- [123] Kaplan M, Ates I, Yüksel M, Ozin YO, Akpinar MY, Topcuoglu C, et al. The Role of Oxidative Stress in the Etiopathogenesis of Gluten-sensitive Enteropathy Disease. *J Med Biochem* 2017;36(3):243–250. doi:10.1515/jomb-2017-0017, PMID:30568541.
- [124] Ruera CN, Miculán E, Pérez F, Ducca G, Carasi P, Chirico FG. Sterile inflammation drives multiple programmed cell death pathways in the gut. *J Leukoc Biol* 2021;109(1):211–221. doi:10.1002/JLB.3MA0820-660R, PMID:32946645.
- [125] Gnodi E, Meneveri R, Barisani D. Celiac disease: From genetics to epigenetics. *World J Gastroenterol* 2022;28(4):449–463. doi:10.3748/wjg.v28.i4.449, PMID:35125829.
- [126] Olazagoitia-Garmendia A, Sebastian-delaCruz M, Castellanos-Rubio A. Involvement of lncRNAs in celiac disease pathogenesis. *Int Rev Cell Mol Biol* 2021;358:241–264. doi:10.1016/bs.ircmb.2020.10.004, PMID:33707056.
- [127] Ferretti G, Baccetti T, Masciangeli S, Saturni L. Celiac disease, inflammation and oxidative damage: a nutrigenetic approach. *Nutrients* 2012;4(4):243–257. doi:10.3390/nu4040243, PMID:22606367.
- [128] Giovannini C, Maiuri L, De Vincenzi M. Cytotoxic effect of prolamin-derived peptides on in vitro cultures of cell line Caco-2: Implications for coeliac disease. *Toxicol In Vitro* 1995;9(3):251–255. doi:10.1016/0887-2333(94)00212-d, PMID:20650086.
- [129] Giovannini C, Mancini E, De Vincenzi M. Inhibition of the cellular metabolism of Caco-2 cells by prolamin peptides from cereals toxic for coeliacs. *Toxicol In Vitro* 1996;10(5):533–538. doi:10.1016/s0887-2333(96)00042-2, PMID:20650234.
- [130] Orlando A, Chimienti G, Pesce V, Fracasso F, Lezza AMS, Russo F. An In Vitro Study on Mitochondrial Compensatory Response Induced by Gliadin Peptides in Caco-2 Cells. *Int J Mol Sci* 2019;20(8):1862. doi:10.3390/ijms20081862, PMID:30991726.
- [131] Lerner A, Neidhöfer S, Matthias T. The Gut Microbiome Feelings of the Brain: A Perspective for Non-Microbiologists. *Microorganisms* 2017;5(4):66. doi:10.3390/microorganisms5040066, PMID:29023380.
- [132] Zelnik N, Pacht A, Obeid R, Lerner A. Range of neurologic disorders in patients with celiac disease. *Pediatrics* 2004;113(6):1672–1676. doi:10.1542/peds.113.6.1672, PMID:15173490.
- [133] Lerner A, Makhlouf BF, Eliakim R. Neurological Manifestations of Celiac Disease in Children and Adults Affiliations Celiac disease and environment View project Neurological Manifestations of Celiac Disease in Children and Adults. *Eur Neurol J* 2012;4:15–20.
- [134] Lerner A, Matthias T. Don't forget the exogenous microbial transglutaminases: it is immunogenic and potentially pathogenic. *AIMS Biophys* 2016;3:546–552. doi:10.3934/biophys.2016.4.546.
- [135] Patel SC, Shreya D, Zamora DL, Patel GS, Grossmann I, Rodriguez K, et al. Celiac Disease, Beyond the Bowel: A Review of Its Neurological Manifestations. *Cureus* 2021;13(12):e20112. doi:10.7759/cureus.20112.
- [136] Shaoul R, Lerner A. Associated autoantibodies in celiac disease. *Autoimmun Rev* 2007;6(8):559–565. doi:10.1016/j.autrev.2007.02.006, PMID:17854749.
- [137] Bressan P, Kramer P. Bread and Other Edible Agents of Mental Disease. *Front Hum Neurosci* 2016;10:130. doi:10.3389/fnhum.2016.00130, PMID:27065833.
- [138] Pruijboom L, de Punder K. The opioid effects of gluten exorphins: asymptomatic celiac disease. *J Health Popul Nutr* 2015;33:24. doi:10.1186/s41043-015-0032-y, PMID:26825414.
- [139] Singh S, Sharma P, Pal N, Kumawat M, Shubham S, Sarma DK, et al. Impact of Environmental Pollutants on Gut Microbiome and Mental Health via the Gut-Brain Axis. *Microorganisms* 2022;10(7):1457. doi:10.3390/microorganisms10071457, PMID:35889175.

- [140] Tran VTA, Lee LP, Cho H. Neuroinflammation in neurodegeneration via microbial infections. *Front Immunol* 2022;13:907804. doi:10.3389/fimmu.2022.907804, PMID:36052093.
- [141] Yelland GW. Gluten-induced cognitive impairment ("brain fog") in coeliac disease. *J Gastroenterol Hepatol* 2017;32(Suppl 1):90–93. doi:10.1111/jgh.13706, PMID:28244662.
- [142] Edwards George JB, Aideyan B, Yates K, Voorhees KN, O'Flynn J, Sweet K, et al. Gluten-induced Neurocognitive Impairment: Results of a Nationwide Study. *J Clin Gastroenterol* 2022;56(7):584–591. doi:10.1097/MCG.0000000000001561, PMID:34049371.
- [143] Makhlof S, Messelmann M, Zaouali J, Mrissa R. Cognitive impairment in celiac disease and non-celiac gluten sensitivity: review of literature on the main cognitive impairments, the imaging and the effect of gluten free diet. *Acta Neurol Belg* 2018;118(1):21–27. doi:10.1007/s13760-017-0870-z, PMID:29247390.
- [144] Lichtwark IT, Newnham ED, Robinson SR, Shepherd SJ, Hosking P, Gibson PR, et al. Cognitive impairment in coeliac disease improves on a gluten-free diet and correlates with histological and serological indices of disease severity. *Aliment Pharmacol Ther* 2014;40(2):160–170. doi:10.1111/apt.12809, PMID:24889390.
- [145] Kristensen VA, Valeur J, Brackmann S, Jahnson J, Brunborg C, Tveito K. Attention deficit and hyperactivity disorder symptoms respond to gluten-free diet in patients with coeliac disease. *Scand J Gastroenterol* 2019;54(5):571–576. doi:10.1080/00365521.2019.1608467, PMID:31050907.
- [146] Obrenovich MEM. Leaky Gut, Leaky Brain? *Microorganisms* 2018; 6(4):107. doi:10.3390/microorganisms6040107, PMID:30340384.
- [147] Lerner A, Matthias T. A Silent or Hypo-symptomatic Disease Can Erupt: Acute Presentations of Celiac Disease. *Int J Celiac Dis* 2017;5:129–132. doi:10.12691/ijcd-5-4-1.
- [148] Guarino M, Gambuti E, Alfano F, Strada A, Ciccocioppo R, Lungaro L, et al. Life-threatening onset of coeliac disease: a case report and literature review. *BMJ Open Gastroenterol* 2020;7(1):e000406. doi:10.1136/bmjgast-2020-000406, PMID:32381744.
- [149] Reinke Y, Zimmer KP, Naim HY. Toxic peptides in Frazer's fraction interact with the actin cytoskeleton and affect the targeting and function of intestinal proteins. *Exp Cell Res* 2009;315(19):3442–3452. doi:10.1016/j.yexcr.2009.06.026, PMID:19576210.
- [150] Stricker S, de Laffolie J, Rudloff S, Komorowski L, Zimmer KP. Intracellular Localization of Microbial Transglutaminase and Its Influence on the Transport of Gliadin in Enterocytes. *J Pediatr Gastroenterol Nutr* 2019;68(3):e43–e50. doi:10.1097/MPG.0000000000002171, PMID:30320664.
- [151] Heyman M, Abed J, Lebreton C, Cerf-Bensussan N. Intestinal permeability in coeliac disease: insight into mechanisms and relevance to pathogenesis. *Gut* 2012;61(9):1355–1364. doi:10.1136/gutnl-2011-300327, PMID:21890812.
- [152] Fasano A. All disease begins in the (leaky) gut: role of zonulin-mediated gut permeability in the pathogenesis of some chronic inflammatory diseases. *F1000Res* 2020;9(F1000 Faculty Rev):69. doi:10.12688/f1000research.20510.1, PMID:32051759.
- [153] Salmi TT, Collin P, Korponay-Szabó IR, Laurila K, Partanen J, Huhtala H, et al. Endomysial antibody-negative coeliac disease: clinical characteristics and intestinal autoantibody deposits. *Gut* 2006;55(12):1746–1753. doi:10.1136/gut.2005.071514, PMID:16571636.
- [154] Lindfors K, Mäki M, Kaukinen K. Transglutaminase 2-targeted autoantibodies in celiac disease: Pathogenetic players in addition to diagnostic tools? *Autoimmun Rev* 2010;9(11):744–749. doi:10.1016/j.autrev.2010.06.003, PMID:20547248.
- [155] Mazumdar K, Alvarez X, Borda JT, Dufour J, Martin E, Bethune MT, et al. Visualization of transepithelial passage of the immunogenic 33-residue peptide from alpha-2 gliadin in gluten-sensitive macaques. *PLoS One* 2010;5(4):e10228. doi:10.1371/journal.pone.0010228, PMID:20419103.
- [156] Bethune MT, Siegel M, Howles-Banerji S, Khosla C. Interferon-gamma released by gluten-stimulated celiac disease-specific intestinal T cells enhances the transepithelial flux of gluten peptides. *J Pharmacol Exp Ther* 2009;329(2):657–668. doi:10.1124/jpet.108.148007, PMID:19218531.
- [157] Lindstad CB, Dewan AE, Stamnaes J, Sollid LM, du Pré MF. TG2-gluten complexes as antigens for gluten-specific and transglutaminase-2 specific B cells in celiac disease. *PLoS One* 2021;16(11):e0259082. doi:10.1371/journal.pone.0259082, PMID:34731200.
- [158] Discepolo V, Lania G, Ten Eikelder MLG, Nanayakkara M, Sepe L, Tufano R, et al. Pediatric Celiac Disease Patients Show Alterations of Dendritic Cell Shape and Actin Rearrangement. *Int J Mol Sci* 2021;22(5):2708. doi:10.3390/ijms22052708, PMID:33800150.
- [159] Upadhyay D, Das P, Dattagupta S, Makharia GK, Jagannathan NR, Sharma U. NMR based metabolic profiling of patients with potential celiac disease elucidating early biochemical changes of gluten-sensitivity: A pilot study. *Clin Chim Acta* 2022;531:291–301. doi:10.1016/j.cca.2022.04.999, PMID:35489390.
- [160] Pennington CL, Dufresne CP, Fanciulli G, Wood TD. Detection of Gluten Exorphin B4 and B5 in Human Blood by Liquid Chromatography-Mass Spectrometry/Mass Spectrometry. *Open Spectrosc J* 2007;1:9–16. doi:10.2174/1874383800701010009.
- [161] Fanciulli G, Pennington CL, Dufresne CP, Wood TD. Gluten exorphins in human blood. *Pharmacol Res* 2020;160:105084. doi:10.1016/j.phrs.2020.105084, PMID:32693107.
- [162] Tan IL, Coutinho da Almeida R, Modderman R, Stachurska A, Deekens J, Barisani D, et al. Circulating miRNAs as Potential Biomarkers for Celiac Disease Development. *Front Immunol* 2021;12:734763. doi:10.3389/fimmu.2021.734763, PMID:34950132.
- [163] Paolini A, Sarshar M, Fellini C, Bruno SP, Rostami-Nejad M, Ferretti F, et al. Biomarkers to Monitor Adherence to Gluten-Free Diet by Celiac Disease Patients: Gluten Immunogenic Peptides and Urinary miRNAs. *Foods* 2022;11(10):1380. doi:10.3390/foods11101380, PMID:35626950.
- [164] Burger JPW, van Lochem EG, Roovers EA, Drenth JPH, Wahab PJ. Dose-Escalating (50-500 mg) Gluten Administration Leads to Detectable Gluten-Immunogenic-Peptides in Urine of Patients with Coeliac Disease Which Is Unrelated to Symptoms, a Placebo Controlled Trial. *Nutrients* 2022;14(9):1771. doi:10.3390/nu14091771, PMID:35565738.
- [165] Monachesi C, Verma AK, Catassi GN, Franceschini E, Gatti S, Gesuita R, et al. Determination of Urinary Gluten Immunogenic Peptides to Assess Adherence to the Gluten-Free Diet: A Randomized, Double-Blind, Controlled Study. *Clin Transl Gastroenterol* 2021;12(10):e00411. doi:10.14309/ctg.0000000000000411, PMID:34613954.
- [166] Palanski BA, Weng N, Zhang L, Hilmer AJ, Fall LA, Swaminathan K, et al. An efficient urine peptidomics workflow identifies chemically defined dietary gluten peptides from patients with celiac disease. *Nat Commun* 2022;13(1):888. doi:10.1038/s41467-022-28353-1, PMID:35173144.
- [167] Piacentini M, D'Eletto M, Farrace MG, Rodolfo C, Del Nonno F, Ippolito G, et al. Characterization of distinct sub-cellular location of transglutaminase type II: changes in intracellular distribution in physiological and pathological states. *Cell Tissue Res* 2014;358(3):793–805. doi:10.1007/s00441-014-1990-x, PMID:25209703.
- [168] Tatsukawa H, Hitomi K. Role of Transglutaminase 2 in Cell Death, Survival, and Fibrosis. *Cells* 2021;10(7):1842. doi:10.3390/cells10071842, PMID:34360011.
- [169] Esposito C, Paparo F, Caputo I, Rossi M, Maglio M, Sblattero D, et al. Anti-tissue transglutaminase antibodies from coeliac patients inhibit transglutaminase activity both in vitro and in situ. *Gut* 2002;51(2):177–181. doi:10.1136/gut.51.2.177, PMID:12117875.
- [170] Hadjivassiliou M, Mäki M, Sanders DS, Williamson CA, Grünewald RA, Woodroffe NM, et al. Autoantibody targeting of brain and intestinal transglutaminase in gluten ataxia. *Neurology* 2006;66(3):373–377. doi:10.1212/01.wnl.0000196480.55601.3a, PMID:16476935.
- [171] Hadjivassiliou M, Aeschlimann P, Sanders DS, Mäki M, Kaukinen K, Grünewald RA, et al. Transglutaminase 6 antibodies in the diagnosis of gluten ataxia. *Neurology* 2013;80(19):1740–1745. doi:10.1212/WNL.0b013e318291070, PMID:23576621.
- [172] Mitoma H, Adhikari K, Aeschlimann D, Chattopadhyay P, Hadjivassiliou M, Hampe CS, et al. Consensus Paper: Neuroimmune Mechanisms of Cerebellar Ataxias. *Cerebellum* 2016;15(2):213–232. doi:10.1007/s12311-015-0664-x, PMID:25823827.
- [173] Boscolo S, Lorenzon A, Sblattero D, Floriani F, Stebel M, Marzari R, et al. Anti transglutaminase antibodies cause ataxia in mice. *PLoS One* 2010;5(3):e9698. doi:10.1371/journal.pone.0009698, PMID:20300628.

- [174] Hadjivassiliou M, Williamson CA, Woodroffe N. The immunology of gluten sensitivity: beyond the gut. *Trends Immunol* 2004;25(11):578–582. doi:10.1016/j.it.2004.08.011, PMID:15489185.
- [175] Giuffrè M, Gazzin S, Zoratti C, Llido JP, Lanza G, Tiribelli C, et al. Celiac Disease and Neurological Manifestations: From Gluten to Neuroinflammation. *Int J Mol Sci* 2022;23(24):15564. doi:10.3390/ijms232415564, PMID:36555205.
- [176] Bashir Y, Khan AU. The interplay between the gut-brain axis and the microbiome: A perspective on psychiatric and neurodegenerative disorders. *Front Neurosci* 2022;16:1030694. doi:10.3389/fnins.2022.1030694, PMID:36389228.
- [177] Lerner A, Matthias T, Aminov R. Potential Effects of Horizontal Gene Exchange in the Human Gut. *Front Immunol* 2017;8:1630. doi:10.3389/fimmu.2017.01630, PMID:29230215.
- [178] Philip A, White ND. Gluten, Inflammation, and Neurodegeneration. *Am J Lifestyle Med* 2022;16(1):32–35. doi:10.1177/15598276211049345, PMID:35185424.
- [179] Lerner A, Jeremias P, Matthias T. Gut-thyroid axis and celiac disease. *Endocr Connect* 2017;6(4):R52–R58. doi:10.1530/EC-17-0021, PMID:28381563.
- [180] Samasca G, Ajay R, Sur D, Aldea C, Sur L, Floca E, et al. Polyautoimmunity - The missing ingredient. *Autoimmun Rev* 2018;17(8):840–841. doi:10.1016/j.autrev.2018.03.008, PMID:29885545.
- [181] Lerner A, Matthias T. Autoimmune Thyroid Diseases in Celiac Disease: If and When to Screen? *Int J Celiac Dis* 2016;4(4):124–126. doi:10.12691/IJCD-4-4-10.
- [182] Liontiris MI, Mazokopakis EE. A concise review of Hashimoto thyroiditis (HT) and the importance of iodine, selenium, vitamin D and gluten on the autoimmunity and dietary management of HT patients. Points that need more investigation. *Hell J Nucl Med* 2017;20(1):51–56. doi:10.1967/s002449910507, PMID:28315909.
- [183] Malandrini S, Trimboli P, Guzzaloni G, Virili C, Lucchini B. What about TSH and Anti-Thyroid Antibodies in Patients with Autoimmune Thyroiditis and Celiac Disease Using a Gluten-Free Diet? A Systematic Review. *Nutrients* 2022;14(8):1681. doi:10.3390/nu14081681, PMID:35458242.
- [184] Duntas LH. Does celiac disease trigger autoimmune thyroiditis? *Nat Rev Endocrinol* 2009;5(4):190–191. doi:10.1038/nrendo.2009.46, PMID:19352315.
- [185] Naeyer AJ, Shah J, Hernandez L, Kim SY, Ciaccio EJ, Cheng J, et al. Tissue transglutaminase antibodies in individuals with celiac disease bind to thyroid follicles and extracellular matrix and may contribute to thyroid dysfunction. *Thyroid* 2008;18(11):1171–1178. doi:10.1089/thy.2008.0110, PMID:19014325.
- [186] Vojdani A, Afar D, Vojdani E. Reaction of Lectin-Specific Antibody with Human Tissue: Possible Contributions to Autoimmunity. *J Immunol Res* 2020;2020:1438957. doi:10.1155/2020/1438957, PMID:32104714.
- [187] Kemppainen E, Salmi T, Lindfors K. Missing Insight Into T and B Cell Responses in Dermatitis Herpetiformis. *Front Immunol* 2021;12:657280. doi:10.3389/fimmu.2021.657280, PMID:33854513.
- [188] Cannistraci C, Lesnoni La Parola I, Cardinali G, Bolasco G, Aspate N, Stigliano V, et al. Co-localization of IgA and TG3 on healthy skin of coeliac patients. *J Eur Acad Dermatol Venereol* 2007;21(4):509–514. doi:10.1111/j.1468-3083.2006.02017.x, PMID:17373979.
- [189] Malkovics T, Koszorú K, Kárpáti S, Arató A, Görög A, Sárdy M. The many-faced gluten sensitivity: Gluten-induced autoimmunity from dermatological point of view. *Orv Hetil* 2021;162(28):1107–1118. doi:10.1556/650.2021.32046, PMID:34252043.
- [190] Goodwin G. Type 1 Diabetes Mellitus and Celiac Disease: Distinct Autoimmune Disorders That Share Common Pathogenic Mechanisms. *Horm Res Paediatr* 2019;92(5):285–292. doi:10.1159/000503142, PMID:31593953.
- [191] Hamilton-Williams EE, Lorca GL, Norris JM, Dunne JL. A Triple Threat? The Role of Diet, Nutrition, and the Microbiota in T1D Pathogenesis. *Front Nutr* 2021;8:600756. doi:10.3389/fnut.2021.600756, PMID:33869260.
- [192] Hakola L, Miettinen ME, Syrjälä E, Åkerlund M, Takkinen HM, Korhonen TE, et al. Association of Cereal, Gluten, and Dietary Fiber Intake With Islet Autoimmunity and Type 1 Diabetes. *JAMA Pediatr* 2019;173(10):953–960. doi:10.1001/jamapediatrics.2019.2564, PMID:31403683.
- [193] Al Theyab A, Almutairi T, Al-Suwaidi AM, Bendriss G, McVeigh C, Chaari A. Epigenetic Effects of Gut Metabolites: Exploring the Path of Dietary Prevention of Type 1 Diabetes. *Front Nutr* 2020;7:563605. doi:10.3389/fnut.2020.563605, PMID:33072796.
- [194] Lund-Blin NA, Tapia G, Mårlid K, Brantsaeter AL, Njølstad PR, Joner G, et al. Maternal and child gluten intake and association with type 1 diabetes: The Norwegian Mother and Child Cohort Study. *PLoS Med* 2020;17(3):e1003032. doi:10.1371/journal.pmed.1003032, PMID:32119659.
- [195] Meijer CR, Discepolo V, Troncone R, Mearin ML. Does infant feeding modulate the manifestation of celiac disease and type 1 diabetes? *Curr Opin Clin Nutr Metab Care* 2017;20(3):222–226. doi:10.1097/MCO.0000000000000367, PMID:28230702.
- [196] Haupt-Jorgensen M, Holm LJ, Josefson K, Buschard K. Possible Prevention of Diabetes with a Gluten-Free Diet. *Nutrients* 2018;10(11):1746. doi:10.3390/nu10111746, PMID:30428550.
- [197] van Lummel M, Duinkerken G, van Veelen PA, de Ru A, Cordfunke R, Zaldumbide A, et al. Posttranslational modification of HLA-DQ binding islet autoantigens in type 1 diabetes. *Diabetes* 2014;63(1):237–247. doi:10.2337/db12-1214, PMID:24089515.
- [198] Füchtenbusch M, Ziegler AG, Hummel M. Elimination of dietary gluten and development of type 1 diabetes in high risk subjects. *Rev Diabet Stud* 2004;1(1):39–41. doi:10.1900/RDS.2004.1.39, PMID:17491663.
- [199] Hansen CHF, Larsen CS, Zachariassen LF, Mentzel CMJ, Laigaard A, Krych L, et al. Gluten-free diet reduces autoimmune diabetes mellitus in mice across multiple generations in a microbiota-independent manner. *J Autoimmun* 2022;127:102795. doi:10.1016/j.jaut.2022.102795, PMID:35101708.
- [200] Haupt-Jorgensen M, Groule V, Reibel J, Buschard K, Pedersen AML. Gluten-free diet modulates inflammation in salivary glands and pancreatic islets. *Oral Dis* 2022;28(3):639–647. doi:10.1111/odi.13775, PMID:33432638.
- [201] Boonpheng B, Cheungpasitporn W, Wijarnpreecha K. Renal disease in patients with celiac disease. *Minerva Med* 2018;109(2):126–140. doi:10.23736/S0026-4806.17.05403-9, PMID:28974086.
- [202] Lerner A, Berthelot L, Jeremias P, Matthias T, Abbad L, Monteiro RC. Gluten, Transglutaminase, Celiac Disease and IgA Nephropathy. *J Clin Cell Immunol* 2017;8:2. doi:10.4172/2155-9899.1000499.
- [203] Abbad L, Monteiro RC, Berthelot L. Food antigens and Transglutaminase 2 in IgA nephropathy: Molecular links between gut and kidney. *Mol Immunol* 2020;121:1–6. doi:10.1016/j.molimm.2020.02.019, PMID:32135400.
- [204] Habura I, Fiedorowicz K, Woźniak A, Idasiak-Piechocka I, Kosikowski P, Oko A. IgA nephropathy associated with coeliac disease. *Cent Eur J Immunol* 2019;44(1):106–108. doi:10.5114/ceji.2019.84021, PMID:31114445.
- [205] Aggarwal M, Garg R, Kumar P, Lindenmeyer CC, Wakim-Fleming J, Jansson-Knodel C, et al. Bi-directional Relationship Between Celiac Disease and Liver Chemistries: A Systematic Review and Meta-Analysis. *Dig Dis Sci* 2023;68(4):1369–1380. doi:10.1007/s10620-022-07663-w, PMID:36002677.
- [206] Rubio-Tapia A, Murray JA. The Liver and Celiac Disease. *Clin Liver Dis* 2019;23(2):167–176. doi:10.1016/j.cld.2018.12.001, PMID:30947869.
- [207] Dutta R, Iqbal A, Das P, Palanichamy JK, Singh A, Mehtab W, et al. Liver involvement in patients with coeliac disease: proof of causality using IgA/anti-TG2 colocalisation techniques. *J Clin Pathol* 2021;74(12):766–773. doi:10.1136/jclinpath-2020-206735, PMID:33789921.
- [208] Afrasiabi R, Siroop PA, Albini SM, Rosenbaum HM, Piscatelli RL. Recurrent pericarditis and dermatitis herpetiformis. Evidence for immune complex deposition in the pericardium. *Chest* 1990;97(4):1006–1007. doi:10.1378/chest.97.4.1006, PMID:2323232.
- [209] Santoro L, De Matteis G, Fuorlo M, Giupponi B, Martone AM, Landi F, et al. Atherosclerosis and cardiovascular involvement in celiac disease: the role of autoimmunity and inflammation. *Eur Rev Med Pharmacol Sci* 2017;21:5437–5444. doi:10.26355/EUR-REV\_201712\_13932.
- [210] Lerner A, Matthias T. Celiac Disease: Intestinal, Heart and Skin Interconnections. *Undefined* 2015;3:28–30. doi:10.12691/IJCD-3-1-6.

- [211] Lerner A. The Gut Feeling of the Heart: Pathophysiological Pathways in the Gut-heart Axis in Celiac Disease. *Int J Celiac Dis* 2020;8(4):120–123. doi:10.12691/IJCD-8-4-2.
- [212] Fousekis FS, Bekar ET, Mitselos IV, Milionis H, Christodoulou DK. Thromboembolic complications and cardiovascular events associated with celiac disease. *Ir J Med Sci* 2021;190(1):133–141. doi:10.1007/s11845-020-02315-2, PMID:32691305.
- [213] Lerner A, Blank M. Hypercoagulability in celiac disease—an update. *Autoimmun Rev* 2014;13(11):1138–1141. doi:10.1016/j.autrev.2014.07.004, PMID:25149392.
- [214] Lerner A, Agmon-Levin N, Shapira Y, Gilburd B, Reuter S, Lavi I, et al. The thrombophilic network of autoantibodies in celiac disease. *BMC Med* 2013;11:89. doi:10.1186/1741-7015-11-89, PMID:23556408.
- [215] Sategna-Guidetti C, Franco E, Martini S, Bobbio M. Binding by serum IgA antibodies from patients with coeliac disease to monkey heart tissue. *Scand J Gastroenterol* 2004;39(6):540–543. doi:10.1080/00365520410008764, PMID:15223677.
- [216] Volta U, Ballardini G, Molinaro N, De Franceschi L, Groff P, Bianchi FB. Specific reactivity of fluorescein isothiocyanate-conjugated separated IgG and IgA from celiac disease sera on human tissues. *Int J Clin Lab Res* 1995;25(2):110–115. doi:10.1007/BF02592367, PMID:7663005.
- [217] Mayer EA, Nance K, Chen S. The Gut-Brain Axis. *Annu Rev Med* 2022;73:439–453. doi:10.1146/annurev-med-042320-014032, PMID:34669431.
- [218] Mohan M, Okeoma CM, Sestak K. Dietary Gluten and Neurodegeneration: A Case for Preclinical Studies. *Int J Mol Sci* 2020;21(15):5407. doi:10.3390/ijms21155407, PMID:32751379.
- [219] Vojdani A, Vojdani E. The Role of Exosomes in the Pathophysiology of Autoimmune Diseases I: Toxic Chemicals and Food. *Pathophysiology* 2021;28(4):513–543. doi:10.3390/pathophysiology28040034, PMID:35366249.
- [220] Vojdani A, Lerner A, Vojdani E. Cross-Reactivity and Sequence Homology Between Alpha-Synuclein and Food Products: A Step Further for Parkinson's Disease Synucleinopathy. *Cells* 2021;10(5):1111. doi:10.3390/cells10051111, PMID:34063062.
- [221] Vojdani A, Kharrazian D, Mukherjee PS. The prevalence of antibodies against wheat and milk proteins in blood donors and their contribution to neuroimmune reactivities. *Nutrients* 2013;6(1):15–36. doi:10.3390/nu6010015, PMID:24451306.
- [222] Vojdani A, O'Bryan T, Green JA, McCandless J, Woeller KN, Vojdani E, et al. Immune response to dietary proteins, gliadin and cerebellar peptides in children with autism. *Nutr Neurosci* 2004;7(3):151–161. doi:10.1080/10284150400004155, PMID:15526989.
- [223] Vojdani A, Vojdani E. Amyloid-Beta 1-42 Cross-Reactive Antibody Prevalent in Human Sera May Contribute to Intraneuronal Deposition of A-Beta-P-42. *Int J Alzheimers Dis* 2018;2018:1672568. doi:10.1155/2018/1672568, PMID:30034864.
- [224] Torsten M, Aaron L. Microbial Transglutaminase Is Immunogenic and Potentially Pathogenic in Pediatric Celiac Disease. *Front Pediatr* 2018;6:389. doi:10.3389/fped.2018.00389, PMID:30619787.
- [225] Aaron L, Torsten M. Microbial transglutaminase: A new potential player in celiac disease. *Clin Immunol* 2019;199:37–43. doi:10.1016/j.clim.2018.12.008, PMID:30543926.
- [226] Lerner A, Matthias T. Microbial Transglutaminase is Beneficial to Food Industries but a Caveat to Public Health. *Med One* 2019;4:e190001. doi:10.20900/mo.20190001.
- [227] Lerner A, Matthias T. Processed Food Additive Microbial Transglutaminase and Its Cross-Linked Gliadin Complexes Are Potential Public Health Concerns in Celiac Disease. *Int J Mol Sci* 2020;21(3):1127. doi:10.3390/ijms21031127, PMID:32046248.
- [228] Lerner A, Matthias T. Microbial transglutaminase should be considered as an environmental inducer of celiac disease. *World J Clin Cases* 2019;7(22):3912–3914. doi:10.12998/wjcc.v7.i22.3912, PMID:31799322.
- [229] Lerner A, Benzvi C. Microbial Transglutaminase Is a Very Frequently Used Food Additive and Is a Potential Inducer of Autoimmune/Neurodegenerative Diseases. *Toxics* 2021;9(10):233. doi:10.3390/toxics9100233, PMID:34678929.
- [230] Lerner A, Ramesh A, Matthias T. Are Non-Celiac Autoimmune Diseases Responsive to Gluten-Free Diet? *Int J Celiac Dis* 2017;5:164–167. doi:10.12691/IJCD-5-4-6.
- [231] Mikulska AA, Karaźniewicz-Łada M, Filipowicz D, Ruchała M, Główka FK. Metabolic Characteristics of Hashimoto's Thyroiditis Patients and the Role of Microelements and Diet in the Disease Management-An Overview. *Int J Mol Sci* 2022;23(12):6580. doi:10.3390/ijms23126580, PMID:35743024.
- [232] Passali M, Josefson K, Frederiksen JL, Antvorskov JC. Current Evidence on the Efficacy of Gluten-Free Diets in Multiple Sclerosis, Psoriasis, Type 1 Diabetes and Autoimmune Thyroid Diseases. *Nutrients* 2020;12(8):2316. doi:10.3390/nu12082316, PMID:32752175.
- [233] Thomsen HL, Jessen EB, Passali M, Frederiksen JL. The role of gluten in multiple sclerosis: A systematic review. *Mult Scler Relat Disord* 2019;27:156–163. doi:10.1016/j.msard.2018.10.019, PMID:30384202.
- [234] Lidón AC, Patricia ML, Vinesh D, Marta MS. Evaluation of Gluten Exclusion for the Improvement of Rheumatoid Arthritis in Adults. *Nutrients* 2022;14(24):5396. doi:10.3390/nu14245396, PMID:36558555.
- [235] Truzzi F, Tibaldi C, Whittaker A, Diloo S, Spisni E, Dinelli G. Pro-Inflammatory Effect of Gliadins and Glutenins Extracted from Different Wheat Cultivars on an In Vitro 3D Intestinal Epithelium Model. *Int J Mol Sci* 2020;22(1):172. doi:10.3390/ijms22010172, PMID:33375311.
- [236] Cai Y, Folkerts J, Folkerts G, Maurer M, Braber S. Microbiota-dependent and -independent effects of dietary fibre on human health. *Br J Pharmacol* 2020;177(6):1363–1381. doi:10.1111/bph.14871, PMID:31663129.
- [237] Tjellström B, Höglberg L, Stenhammar L, Fälth-Magnusson K, Magnusson KE, Norin E, et al. Faecal short-chain fatty acid pattern in childhood coeliac disease is normalised after more than one year's gluten-free diet. *Microb Ecol Health Dis* 2013;24(1):20905. doi:10.3402/mehd.v24i0.20905, PMID:24082880.
- [238] Pynnönen PA, Isometsä ET, Verkasalo MA, Kähkönen SA, Sipilä I, Savilahti E, et al. Gluten-free diet may alleviate depressive and behavioural symptoms in adolescents with coeliac disease: a prospective follow-up case-series study. *BMC Psychiatry* 2005;5:14. doi:10.1186/1471-244X-5-14, PMID:15774013.
- [239] Addolorato G, Capristo E, Ghittoni G, Valeri C, Mascianà R, Accuna C, et al. Anxiety but not depression decreases in coeliac patients after one-year gluten-free diet: a longitudinal study. *Scand J Gastroenterol* 2001;36(5):502–506. doi:10.1080/00365520119754, PMID:11346203.
- [240] Rostami-Nejad M, Taraghikhah N, Ciacci C, Pourhoseingholi MA, Barzegar F, Rezaei-Tavirani M, et al. Anxiety Symptoms in Adult Celiac Patients and the Effect of a Gluten-Free Diet: An Iranian Nationwide Study. *Inflamm Intest Dis* 2020;5(1):42–47. doi:10.1159/000505657, PMID:32232054.
- [241] Reinhorn IM, Bernstein CN, Graff LA, Patten SB, Sareen J, Fisk JD, et al. Social phobia in immune-mediated inflammatory diseases. *J Psychosom Res* 2020;128:109890. doi:10.1016/j.jpsychores.2019.109890, PMID:31816595.
- [242] Euesden J, Danese A, Lewis CM, Maughan B. A bidirectional relationship between depression and the autoimmune disorders - New perspectives from the National Child Development Study. *PLoS One* 2017;12(3):e0173015. doi:10.1371/journal.pone.0173015, PMID:28264010.
- [243] Odineal DD, Gershwin ME. The Epidemiology and Clinical Manifestations of Autoimmunity in Selective IgA Deficiency. *Clin Rev Allergy Immunol* 2020;58(1):107–133. doi:10.1007/s12016-019-08756-7, PMID:31267472.
- [244] Lerner A, Neidhöfer S, Matthias T. The gut-gut axis: Cohabitation of celiac, Crohn's disease and IgA deficiency. *Int J Celiac Dis* 2016;4:68–70. doi:10.12691/IJCD-4-2-1.
- [245] Tan J, McKenzie C, Vuillermin PJ, Goverse G, Vinuesa CG, Meibius RE, et al. Dietary Fiber and Bacterial SCFA Enhance Oral Tolerance and Protect against Food Allergy through Diverse Cellular Pathways. *Cell Rep* 2016;15(12):2809–2824. doi:10.1016/j.celrep.2016.05.047, PMID:27332875.
- [246] Drabińska N, Jarocka-Cyrtka E, Markiewicz LH, Krupa-Kozak U. The Effect of Oligofructose-Enriched Inulin on Faecal Bacterial Counts and Microbiota-Associated Characteristics in Celiac Disease Children Following a Gluten-Free Diet: Results of a Randomized, Placebo-Con-

- trolled Trial. *Nutrients* 2018;10(2):201. doi:10.3390/nu10020201, PMID:29439526.
- [247] Dicks LMT, Vermeulen W. Do Bacteria Provide an Alternative to Cancer Treatment and What Role Does Lactic Acid Bacteria Play? *Microorganisms* 2022;10(9):1733. doi:10.3390/microorganisms10091733, PMID:36144335.
- [248] Liu P, Wang Y, Yang G, Zhang Q, Meng L, Xin Y, et al. The role of short-chain fatty acids in intestinal barrier function, inflammation, oxidative stress, and colonic carcinogenesis. *Pharmacol Res* 2021;165:105420. doi:10.1016/j.phrs.2021.105420, PMID:33434620.
- [249] Casas R, Sacanella E, Estruch R. The immune protective effect of the Mediterranean diet against chronic low-grade inflammatory diseases. *Endocr Metab Immune Disord Drug Targets* 2014;14(4):245–254. doi:10.2174/1871530314666140922153350, PMID:25244229.
- [250] Spyridaki A, Psylinakis E, Chatzivasili D, Thalassinos N, Kounelaki V, Charonitaki A, et al. Adherence to the Mediterranean diet is linked to reduced psychopathology in female celiac disease patients. *Psychol Health Med* 2023;28(6):1634–1639. doi:10.1080/13548506.2022.2052329, PMID:35282714.
- [251] Morreale F, Agnoli C, Roncoroni L, Sieri S, Lombardo V, Mazzeo T, et al. Are the dietary habits of treated individuals with celiac disease adherent to a Mediterranean diet? *Nutr Metab Cardiovasc Dis* 2018;28(11):1148–1154. doi:10.1016/j.numecd.2018.06.021, PMID:30143412.
- [252] Lerner A, Benzvi C, Vojdani A. Cross-reactivity and sequence similarity between microbial transglutaminase and human tissue antigens. *Sci Rep* 2023;13(1):17526. doi:10.1038/s41598-023-44452-5, PMID:37845267.
- [253] Bischoff SC, Barbara G, Buurman W, Ockhuizen T, Schulzke JD, Serino M, et al. Intestinal permeability—a new target for disease prevention and therapy. *BMC Gastroenterol* 2014;14:189. doi:10.1186/s12876-014-0189-7, PMID:25407511.
- [254] Barone MV, Salvatore A. Pro-Inflammatory Nutrient: Focus on Gladin and Celiac Disease. *Int J Mol Sci* 2022;23(10):5577. doi:10.3390/ijms23105577, PMID:35628388.
- [255] English J, Connolly L, Stewart LD. Increased Intestinal Permeability: An Avenue for the Development of Autoimmune Disease? Springer Netherlands 2024;16:575–605. doi:10.1007/s12403-023-00578-5.
- [256] Guandalini S. Editorial: Gluten: yes, no, maybe. *Front Med (Lausanne)* 2023;10:1225139. doi:10.3389/fmed.2023.1225139, PMID:37359016.